Therapeutic Efficacy of *Thymus serpyllum* Derived Silver Nanoparticles in Streptozotocin Induced Diabetic BALB/c Mice



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I, Ayesha Imran, declare that this research work titled "Therapeutic Efficacy of *Thymus serpyllum* Derived Silver Nanoparticles in Streptozotocin Induced Diabetic BALB/c Mice" is my own work. The work has not been presented elsewhere for assessment. The work here in was carried out while I was a post-graduate student at Atta-ur-Rahman School of Applied Biosciences, NUST under the supervision of Dr. Peter John. The material that has been used from other sources has been properly acknowledged/ referred.

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LIST OF ACRONYMS

AgNPs	Silver Nanoparticles
AgNO3	Silver nitrate
μg	Microgram
2D	2 Dimensional
3D	3 dimensional
ANOVA	Analysis of Variance
GLUT4	Glucose Transporter 4
GLUT2	Glucose Transporter 2
IRS2	Insulin Receptor Substrate 2
INS2	Insulin 2
АМРК	Adenosine Monophosphate Kinase
T2DM	Tpe2 Diabetes Mellitus
T1DM	Tpe1 Diabetes Mellitus
STZ	Streptozotocin
UV Vis	Ultraviolet Visible
XRD	X-ray Dispersive Spectroscopy
FBG	Fasting Blood Glucose level
PDB	Protein Data Bank

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ABSTRACT

Type 2 diabetes mellitus (DM) is a chronic metabolic disorder and is characterized by means of consistent hyperglycemia as a consequence of either entire or incomplete deficiency of insulin secretion and action of insulin. The most commonly used therapeutic agent used for type 2 diabetes mellitus is metformin. However, it has various side effects which include its slow mode of action and gastrointestinal infections. Therefore scientists are trying to overcome these limitations by promoting nanomedicine that can be synthesized from biological sources. In nanomedicine, silver nanoparticles have gained more importance due to their strong anti-diabetic, anti-oxidant and antimicrobial properties.

The present study reports a green, economical and biological means for the efficient production of the silver nanoparticles (AgNPs) from the extract of *Thymus serpyllum*. This synthesis method was also optimized under different physiochemical parameters such as temperature and molar concentration ratios to obtain narrow size distribution of AgNPs. XRD was used to characterize these silver nanoparticles. To induce diabetes in BALB/c mice, streptozotocin (STZ) injections were given along with high fat diet. Primers for Glut4, INS2, IRS2 were designed by Primer 3 Plus and then validated by UCSC In-Silico PCR.

The results were initially assessed by UV-VIS spectroscopy, showing maximum absorbance at the characteristic wavelength of 400nm. The shape of these silver nanoparticles was found spherical. XRD pattern of silver nanoparticles showed six intensive peaks. The difference between Fasting Blood Glucose (FBG) level and body weight of control and experimental mice showed the successful induction of diabetes in BALB/c mice. UCSC In-Silico PCR gives the primers that showed binding to the target region and showed no off-target binding. Thus, silver nanoparticles having anti-diabetic potential can be synthesized from *Thymus serpyllum* extract under optimized conditions.

CHAPTER 1

INTRODUCTION

1. Diabetes mellitus

Diabetes mellitus is delineated through means of persistent hyperglycemia as well as deterioration in the metabolism of carbohydrates, lipids, proteins that is imputable to either absolute or incomplete deficiency of insulin secretion or insulin action. The persistent hyperglycemia of the diabetes is interconnected with protracted damage, pathology, anomaly, and malfunction of various organs, substantially the eyes, kidneys, nerves, heart, and blood vessels. There are two crucial forms of diabetes, Type1 Diabetes Mellitus, T1DM and Type2 Diabetes Mellitus, T2DM (Galtier, 2010; Diabetes Care 2014).

1.1 Type2 Diabetes Mellitus

Type2 Diabetes Mellitus is additionally known as **Insulin Independent Diabetes Mellitus** (IDDM). It is an inimitable category of DM that corresponds about 90% to 95% of whole diabetic inmates (Tripathi, 2006). It is inevitable that the cases will be increase to 439 million by 2030 (Chen et al., 2011). T2DM is a **polygenic disorder** delineated by various inadequacies in insulin activity within internal organs and the scarcities in pancreatic secretion of insulin, a fact which ultimately stimulates failure insulin secreting cells in pancreas (Israili, 2011). Type2 DM occurs as a result of interactions between genetic, behavioral as well as environmental risk factors (Chen, 2011).

T2DM inpatients can further vulnerable to diverse sorts of complications i.e. transitory and protracted time period complications. The complicacies (figure 1.1) consist of **macrovascular**

problems (hypertension, heart attacks, hyperlipidemia, strokes, coronary disorders, cerebral and peripheral vascular abnormalities), **microvascular** problems (retinopathy, neuropathy, and nephropathy) and cancers (Yanling et al., 2014).



Figure 1.1: Complication of Type2 Diabetes Mellitus (Yanling et al., 2014)

Cardiovascular disorder is the crucial incentive of fatality and morbidness in prediabetes as well as T2DM, for which oxidative stress is effectual and influential mechanism (Chaturved, 2007). Diabetic neuropathy can additionally established a correspondence among foot ulcers, amputations, noxious skin injuries and sexual abnormalities. The neuropathy will result into failure of defensive sensibility in the foot that utmost terminates in callous formulation, contusion as well as auxiliary impairment and could additionally bring about contamination in the epidermis (Sanghera DK, Blackett PR, *2012*). Epidemiologic data has established that diabetes can enhance the contingence of cancer for instance colorectal cancer, , bladder

cancer, liver cancer, breast cancer, kidney cancer, that deteriorates relying on sub sites of distinctive cancers (Donadon et al., 2008; Larsson and Wolk, 2011).

1.2 Epidemiology/Prevalence of Diabetes Mellitus

Diabetes is a most important health issue as it has progressed to the petrifying levels. In 2019, there are almost semi quadrillion people (9.3% of adult's 20–79 years) that have diabetes across the board. The predictable numeral of personages (20–79 years) that are suffering from diabetes has been increased by 62% during the period of last 10 years; from 285 million in 2009 to 463 million, at present and will intensified to 10.2% (578 million) by 2030 and 10.9% (700 million) by 2045. The prevalence of diabetes by age and gender in is given in figure 1.2 (Saeedi et al., 2019).



Figure 1.2: Prevalence of diabetes (Saeedi et al., 2019)

The incidence of diabetes has been expanding rapidly and this is the foremost incentive of presbyopia, retrogression of kidneys, stroke, acute myocardial infarction as well as inferior forelimb disjuncture. In 2016, there was a probability that almost 1.6 million deaths were directly caused by diabetes. According to World Health Organization (WHO), diabetes is the seventh leading cause of death in 2016 (International Diabetes Federation, 2017).

Approximately, semi percent (50.1%) of the population/inhabitants accompanied by diabetes do not comprehend that they are suffering from diabetes. Type 2 diabetes depicts the most prevalent category of diabetes. It is responsible for about 90% of entire diabetic inpatients as compare to type 1 diabetes that accounts for approximately 10% (Sarwar et al., 2010). The pervasiveness of type 2 diabetes and prediabetes has increased in Pakistan. Pakistan has approximately 74 lac adults with diabetes and therefore ranked as 10th because of highest number of diabetes cases (Sherin, 2015).

1.3 Pathophysiology/Pathogenesis of Diabetes Mellitus

1.3.1 Normal Glucose Homeostasis

Glucose is transported in the blood to certain tissues to be used as energy. **Insulin** that helps the absorption of glucose originating at blood circulation with the aid of fat tissue and skeletal muscles, is itself symphonized as well as originated by Beta cells that are present inside the pancreas. Insulin serves as the hormone (peptidal hormone) which has phenomenal ability to regulate the assimilation of fat as well as carbohydrate within an individual's body (AlSaraj, 2015).

There are four transporters of glucose (GLUT1, GLUT2, GLUT3 and GLUT4). After intake of meal, blood glucose enters amongst the cell which is facilitated by Glucose transporter type 2 (GLUT2) membrane transporters as shown in the figure 1.3



Figure 1.3.1: Normal Glucose Homeostasis (Arthur, 2006)

The subsequent glucose within the cells is then phosphorylated via means of glucose-6phosphatase toward glucose-6-phosphate which will ultimately enter into glycolysis series, thus triggering pyruvate in addition to adenosine triphosphate (ATP). Pyruvate then intrudes into next phase i.e. tricarboxylic acid (TCA) cycle, as a result of which further ATP will generate. KATP channel will block due to increase in ATP: ADP ratio, which leads to the depolarization of membrane and hence permitting Ca 2+ ions towards the cell facilitated by voltage dependent calcium channel. These ions will then eventually exhilarate the release of insulin (peptidal hormone); Insulin will then bind to the receptors of insulin which will ultimately initiates the phosphorylation based signaling of cascade. For uptake of glucose molecules, the translocation of intracellular GLUT4 to membrane will occur (Lee and Halter, 2017; Arthur, 2006).

1.3.2 Pathophysiology of Type2 Diabetes Mellitus

Impaired secretion of insulin and insulin resistance make a contribution towards the development of pathophysiological conditions. Various organs participate during the physiological progression of type2 diabetes (figure 1.3.1). Interruption in the connection among various organs i.e. pancreas, liver, adipose tissue, gut and central nervous system direct towards modification in normal glucose metabolism and type2 diabetes (DeFronzo, 1988; Reaven, 1995)



Figure 1.3.2: Role of organs in pathogenesis of type2 diabetes (DeFronzo, 1988; Reaven, 1995)

1.3.2.1 Impaired insulin secretion

Impaired secretion of insulin will results in decrease in responsiveness of glucose, as it has been observed earlier than pathological progression carried out by complication. Further distinctively, impaired glucose tolerance (IGT) is driven via reduction within initial-phase insulin secretion, which is glucose-responsive. And therefore, a reduction in supplementary secretion of insulin occurs after meals. This will ultimately results postprandial hyperglycemia. Impaired secretion of insulin is generally progressive, and its development includes toxicity of glucose as well as lipo-toxicity. When it will remain untreated, this leads to the reduction of pancreatic cell (Ghani et al., 2008). The procession regarding effective decline relating to pancreatic cell utility consequently causes the permanent blood glucose elevation (figure 1.3.2.1).



Figure 1.3.2.1: Development of type2 diabetes from pancreatic cells (Kaku, 2010)

1.3.2.2 Insulin resistance

This is the critical situation of the type that insulin is not able to perform its function into the body. This deprivation in the efficiency of insulin within predominant organs i.e. liver and muscles is one of the frequent procession characteristic of type 2 diabetes. The subatomic process for the activity of insulin has shown that this complication of resistance in insulin is associated in conformity with inherited factors along with environmental elements (hyperglycemia, free fatty acids, inflammatory mechanism, etc.) (Kaku, 2010).

There are three potent mechanisms for the control of metabolism of glucose. These mechanisms are present in skeletal muscles. This includes the foremost insulin stimulated

glucose transporter GLUT4, hexokinase and glycogen synthase. Consequently, malfunction in skeletal muscles glycogen production plays a chief role in the origination and development of processes involved in Insulin resistance. Similarly, the reduced capability of typical functioning of the above mentioned mechanisms that helps in circulation of insulin is major fundamental for the progression in insulin resistance (Petersen, 2002).

Genetic factor involves not solely polymorphisms in insulin receptor and insulin receptor substrate (IRS)-1 gene that has the ability to influence the signals of insulin. The polymorphisms of thrifty genes such i.e. 3 adrenergic receptor genes as well as uncoupling protein (UCP) gene are related to the genetic factors. These polymorphisms are related with visceral obesity and develop insulin resistance (Matsuda and DeFronzo, 1999).

1.4 Etiology of Type2 Diabetes Mellitus

The origin of T1DM is ultimate. However, T2DM do not have an ultimate and legitimate cause i.e., type2 diabetes mellitus is multifactorial (figure 1.4). The frequencies of development of abnormalities along with its related complexities are dissimilar in the inmates suffering from type2 diabetes. Furthermore, numerous determinants are present which somehow implicated in insulin resistance and other complications (Ismail et al., 2010; Thomas and Philipson, 2015; Zoungs et al., 2009; DeFronzo, 2015).

Type2 diabetes mellitus which is complicated multifactorial ailment, also involves multitudinous genes that are geographically dispersed amongst specific chromosomal regions. It is also related by means of distinctive pathways i.e. metabolic along with cellular signaling mechanisms. The possibility of developing Type2 diabetes can also be associated with dietary as well as environmental constituents. On the other hand, hereditary factors can also participate as a considerable factor for the elucidation of Type2 diabetes (Kaku, 2010)



Figure 1.4: Etiology and pathophysiology of type2 diabetes (DeFronzo, 2015)

1.4.1 Genetic factors

Genome wide association studies (GWAS) has pinpointed 70 loci that are associated among type2 diabetes in range of populations. Stumvoll et al in 2015 had given the substantiality about chances to develop type2 diabetes if the family has positive history of diabetes. This depiction imparts about 2.5 times foremost possibility of developing Type2 diabetes. The foremost pedigree of Type2 diabetes inpatients has almost 15-25% chance to initiate malfunctioned glucose tolerance. This will ultimately lead the procession of Type2 diabetes. In 1995, Pierce et al., illustrated that possibility of the procession and progression of Type2 diabetes is approximately 38-40%, if only a mother or only a father are suffering from Type2 diabetes (Pierce et al., 1995). Tattersal and Fajans in 1975 demonstrated that approximately above 50% chances that the children will develop diabetes, if together mother and father both are suffering from diabetes (Tattersal and Fajans, 1975).

IRS2 gene encodes a protein that has the capability to act as an adaptor to insulin receptor. Here it activates cascade of kinase in pathways i.e. insulin signaling. However in case of type2 diabetes, the resistance in insulin secretion will occur due to suppression of *IRS2* gene. Thus IRS2 gene plays a major role in normal insulin signaling pathway (Zhang and Sun, 2009). In the previous years, it was observed that hyperglycaemia causes some major changes especially in the nervous system by modifying the function of IRS proteins. This ultimately leads to the impairment in diabetic marginal neuropathy as well as insulin signaling mechanism and cascade (Manu, Rachana and Advirao, 2017). In the latest findings, it is determined that polymorphism in Gly1057Asp of IRS-2 gene is mainly associated with T2DM (Ay et al., 2018).

1.4.2 Epigenetic Factors

Viral infections and toxins have the ability to modify the regular metabolism that influence for the epigenetic changes (figure 1.4.2). These modifications have the ability to manipulate the genes. This occurs as a result of two main processes i.e. by developing resistance and by the increasing the susceptibility to disease complications through allele polymorphism (Desiderio et al., 2016). Dietary factors have the ability to modulate epigenetics when an individual has sedentary lifestyle, has large intake of fatty foods and less intake of healthy food (Etchegaray and Mostoslavsky, 2016).

Methylation of DNA is determined as one of the main factor for the initiation of Type2 diabetes that affects secretion as well as sensitivity of insulin. Zhou and his co workers had illustrated almost 17 diverse genes. All these genes are found to be associated with development of type2 diabetes caused by epigenetic factors (Zhou, Sun, Li, & Zhu, 2018). Among all these genes, two genes i.e. insulin present in pancreas and metformin transporter genes that are present in liver are determined to be susceptible for demethylation.

The microvascular complexities of Type2 diabetes is related with the mitochondrial DNA. Methylation of this DNA takes place which ultimately cause various complications (Rorbach-Dolata, Kubis and Piwowar, 2017). Arginine and lysine amino acids present in the histone proteins are acetylated which is one of the factors for procession of type2 diabetes. This Acetylation is considered as an important modification in epigenetics (Rorbach-Dolata, Kubis and Piwowar, 2017).



Figure 1.4.2: Classification of Epigenetic Factors

1.4.3 Ecological Factors

Increase in age along with weight, inadequate consumption of energy, alcohol intake, smoking, etc. are equitable threat elements that leads to the pathogenesis of type2 diabetes mellitus (figure 1.4.3) (Hu et al., 2001) (Belkina and Denis, 2010). **Obesity** (mainly visceral fat obesity) because of deficiency of exercise followed via the reduction in muscle mass, also culminates in insulin resistance, and also closely related to the immediate increase in the magnitude of patients with ages 30 and above (Zimmet et al., 2001). Even mild obesity (weight problems) increases the risk of developing diabetes, approximately 4- to 5-fold.

World Health Organization (WHO) in 2011 illustrated that around 90% of inpatients had developed Type2 diabetes as a result of excess body weight (WHO, 2011).



Figure 1.4.3: Contribution of environmental factors in type2 diabetes (Scheen, 2001)

The modifications in the **nutritional energy resources**, especially the proliferation in intake of fat, reduce intake of starch along with increment in the consumption of plain sugars and reduction in intake of dietary fiber, make a contribution toward obesity as well as initiate deterioration of glucose tolerance (Schulze et al., 2004; Dhingra et al. 2007).

1.4.4 Inflammatory factors

Oxidative stress culminates in the assembly of **reactive oxygen species** (**ROS**) together with other **pro-inflammatory cytokines** and **chemokines** in the beta cells. This will interrupt the blood circulation within the pancreatic cells thus destroying their utility (Ehses et al., 2007; Dedon and Tannenbaum, 2004).

According to different studies, **IL-6** as well as various other inflammatory cytokines has the capability to induce apoptosis in pancreatic islets. They can also act as a predictor and pathogenic marker for the development of T2DM (Pradhan, 2001; Akash et al., 2012). **TNF-alpha** creates an interrelation amongst complications in insulin secretion, obesity and islet hypersensitivity. Therefore, it is considered to play an essential part in the procession of Type2 diabetes (Tilg and Moschen, 2008). Its overproduction among adipose tissue appears to cause the hypersensitivity and beta cell loss present in pancreatic islets. It also produces extra complications in insulin secretion in the marginal tissues (Rosenvinge et al., 2007; Ruan, 2002).

1.5 Symptoms of Diabetes

If type2 diabetes proceeds without treatment, glucose levels in the circulation continue to remain elevated every time. However, this isn't always obvious at first. Type2 diabetes may extend progressively above several years devoid of any evident symptoms. Glucose levels in the blood which remains excessive all the time can initiate the subsequent symptoms: **Extreme thirst, frequent urination, tiredness, nausea along with dizziness**. If an individual has extremely high blood sugar levels, he may experience burdens' and drowsiness and ultimately he will lose consciousness (diabetic coma) (American Diabetes Association, 2017; Yan et al., 2016).

1.6 Diagnostic tests

Diabetes can be diagnosed by the hemoglobin A1C criteria or plasma glucose absorption (fasting or 2-hour plasma glucose) as shown in table 1.6.

1.6.1 Two-Hour Oral Glucose Tolerance Test (OGTT)

In this diagnosis, the plasma glucose level is computed before 2 hours of intake of glucose and 2 hours after the intake of approximately 75-80 gm of glucose. The individual is considered diabetic if glucose level is more than 200 mg/dL in sample taken after 2 hours of glucose intake

1.6.2 Fasting Plasma Glucose (FPG)

A blood sample is withdrawn from an individual after overnight fasting for about 8 hours. As per ADA, if the fasting glucose level is more than 126 mg/dL than the individual is considered diabetic.

1.6.3 Glycated Hemoglobin (Hb) A1C

This test provides an estimate of average blood glucose level in the previous 2 to 3 months of an individual. Patients in which Hb A1C is more than 6.5% are considered as diabetic. it measures the thickness of sugar bound to haemoglobin in red blood cells. The thicker sugar coat on red blood cells indicates the higher level of glucose in blood (Martinez et al., 2019; Lai et al., 2019).

Diagnostic Test	Normal	Diabetic
2-Hour Post Glucose Intake	≤140 mg/dL	≥200 mg/dL
Fating Plasma Glucose	70-99mg/dL	$\geq 126 \text{ mg/dL}$
Glycated Hemoglobin	4-6%	≥6.5%

Table 1.6: Diagnostic criteria of Diabetes

1.7 Treatment of type 2 diabetes

1.7.1 Diet Therapy

Almost 50% cases of type2 diabetes are due to elevated blood sugar levels accompanied by overweight and sedentary lifestyle. Therefore, the aim to decrease the weight of diabetic individual can come under this diet treatment (Wing, 1995; Henry and Gumbiner, 1991). Food items rich in carbohydrate while having least lipid content result in improved glycaemic control and reduction in low density lipids (Franz et al., 1994)

1.7.2 Exercise

In the previous studies (Helmrich et al., 1994; Martin and Wahren, 1993) it is illustrated that exercise has numerous advantages in the anticipation of progression of type 2 diabetes mellitus. It also has the ability to increase sensitivity of glucose and thus to improve levels. Reduced intraabdominal fat, increased activity of glucose transporters (GLUT-4) that are sensitive to insulin present in muscle, improved blood circulation in tissues that are sensitive to insulin, are all the mechanisms through which sensitivity of insulin is restored by exercise (Erisonn et al., 1997)

1.7.3 Pharmacotherapy

The curative agents accessible for treatment of type2 diabetes consist of sulfonylureas and related compounds, biguanides, thiazolidenediones, α -glucosidase inhibitors. Pharmaceutical agents that can act through diverse mechanisms of action must be selected in order to improve glucose values and at the same time minimizing unfavorable effects. Some of the therapeutic agents are enlisted in the table 1.7.3 (Wu et al., 2014).

Class	Drug	Target	Disadvantage
Biguanides	Metformin	Amp-Kinase	Vitamin B12 deficiencyGastrointestinal problems
α-glucosidase inhibitors	Acarbose/ Miglitol/ Voglibose	α-glucosidase	• Gastrointestinal problems
Thiazolidinediones	Troglitazone/ Pioglitazone	PPAR-Υ	Blood cancer riskWeight gain

Table 1.7.3: Therapeutic Agents (Wu et al., 2014)

1.8 Insulin Therapy

Insulin therapy can be used for treatment of type2 diabetes as foremost remedy of extreme hyperglycemia along with additional critical hyperglycemic circumstances. Insulin can be used along with other treatments and combinations for curing type2 diabetes. Burge and Schade illustrated large numbers of latest analogs of insulin that are in pharmaceutical trials (Burge, Schade, 1997). Lispro insulin is the foremost accessible analog of insulin. It represents a two amino acid alteration as compared to typical insulin present in humans (Holleman F, Hoekstra, 1997).

1.9 Antioxidant therapy

The hindrance of intracellular free radical development would present a therapeutic approach in order to avoid oxidative stress and the interrelated diabetic vascular problems. Antioxidants may additionally proceed at diverse levels, hindering the development of ROS or scavenge free radicals, and enhancing the antioxidants defense enzyme performances (Jakus, 2000).

Medicinal floras having antioxidant properties have the ability to be defensive in diabetic rats. This ability is due to scavenging free radicals especially oxygen radicals along with reduction in expressions of molecule-1 protein intercellularly (Ghamarian et al., 2012; Rafieian and Baradaran, 2013; Nasri and Rafieian, 2013). Medicinal floras bear an extensive history for curing the complications. In conventional medicinal treatment it involves almost 800 plants that have the ability to cure diabetes (Rafieian and Baradaran, 2013; Hung et al., 2013; Nasri et al., 2014).

A probable assistance about vegetables and fruit is because of antioxidant components that are present inside them. Therefore, they made their contribution for the suppression of oxidative stress (Hulbert et al., 2005). Vegetables as well as fruits exhibited elevated accumulation of antioxidants, that has the ability to decrease the possibility of developing diabetes especially type2 diabetes (Tamadon et al., 2013).

Plants encompass to a great extent of enchantment due to their defensive along with curative characteristics for the diseases that are very difficult to cure which involves memory deterioration, diabetes, cancer along with heart diseases and many more, all these properties are due to their antioxidant potentials (Taghikhani et al., 2014; Beladi et al., 2014).
1.9.1 Nanoantioxidants

Many plant mediated nanoparticles are potent source of antioxidants that act to reduce the oxidative stress produced as a metabolic response or induced exogenously (Garhwal, 2010). Metallic nanoparticles (NPs) such as magnetic, silver, and gold NPs, nanosized entities that have dimension between 1–100 nm, has the potential to treat type2 diabetes because of their physical along with chemical characteristics and their capability to transform the level of oxidative stress (Lushchak et al., 2018).

Silver nanoparticles have the ability to restore the normal **glucose**, serum **insulin** levels and **glucokinase** activity. It has also the potential to reduce lipid profile by suppressing the oxidative stress and elevating the antioxidant defense system (Shaheen et al., 2016).

1.10 Thymus serpyllum

Thymus serpyllum (wild thyme) associated with the family of Lamiaceae and as mentioned by the World Checklist, constitutes 7534 species, which includes the genus *Thymus* L. having 220 kind of species (Harley et al., 2004).

It is a necessary herbal plant with ethno-botanical importance in diverse regions of the World, peculiarly in Gilgit Baltistan region of Pakistan. Wild thyme for the most part repeatedly intended for treatment of sickness and complications interrelated with gastrointestinal along with respiratory systems (Menkovic et al., 2011; Benitez at al., 2016). *Thymus serpyllum* is considered an imperative supply of remedial agents having **antioxidant**, **antimicrobial**, **antitumor properties**. Wild thyme has various important properties due to which it can be exploited in various industries i.e. medicinal, food, cosmetics and many more (figure 1.10) (Jaric et al., 2015).



Figure 1.10: *Thymus serpyllum* and its properties (Jarić et al., 2015)

1.11 Objectives

- Docking of Silver with Glut 4
- Synthesis of cost effective, nontoxic environment friendly *Thymus serpyllum* mediated silver nanoparticles.
- Characterization of the synthesized biogenic silver nanoparticles by UV-Visible Absorption Spectroscopy and X-Ray Diffraction
- Induction of diabetes mellitus in BALB/C mice.
- Primer designing of INS2, IRS2, Glut4 and β-actin by primer 3 plus and validation by UCSC In-Silico PCR.

CHAPTER 2

LITERATURE REVIEW

2.1 Introduction to Nanotechnology

Nanotechnology is the art about manipulating matter considered as the billionths of meters and nanometer, approximately the mass of 2 or 3 atoms (Kaira and Singh, 2012). Nanotechnology has provided absolutely innovative concepts and innovative approaches/processes in medicinal drug as well as in dentistry which will facilitate us in making superior, former and sure prognosis and to treat patients with least feasible interventions and without any unfavorable outcomes (Tevatia et al., 2016).

On 29th December 1959, the deceased Nobel Prize–winning physicist Richard P. Feynman published a discussion at Caltech on a title "There's Plenty of Room at the Bottom" (Feynman, 1960) at the yearly conference, organized by American Physical Society. He recommended that certain nanomachines, nano-robots as well as nano-devices eventually might be exploit to build up the comprehensive array of atomically defined microscopic equipments, technical as well as assembling materials. Thus the revelation of nanotechnology was evolved (Asiyanbola and Soboyejo, 2008) (Tevatia, 2016). The word "nanotechnology" was described by Tokyo Science University Professor Norio Taniguchi in 1974 paper. Norio defined this term as follows: "Nano-technology' primarily comprises of the processing, dispensation, separation, consolidation, as well as deformation of materials and substances by solitary atom or by solitary molecule" (Taniguchi, 1974). "Engines of Creation: The Coming Era of Nanotechnology" by Dr. K. Eric Drexler is considered as the first book on the subject of nanotechnology Initiative was inaugurated in order to synchronize with the Federal nanotechnology research, progression and development (Ratner, 2002).

2.2 Nanotechnology in medicine (Nanomedicine)

The appliance of nanotechnology in favor of therapeutics, prognosis, examination, as well as inhibition of various complications and diseases is called "nanomedicine". In 1965, the first illustration regrading lipid vesicles by Bangham and his co-workers was presented, that was later on become acknowledged by the term liposomes (Bangham et al., 1965); In 1976 the primary managed dispensed polymer structures related to macromolecules were discussed and demonstrated by Langer and Folkman (Langer and Folkman, 1976); In, 1994, the initial elongated diffusing sheath of nanoparticles made up of polymers was once illustrated by Gref and his co-workers (Gref et al., 1994); In 1998 the foremost quantum dot produced as result of bioconjugation was discussed and illustrated by Bruchez and his co-workers (Bruchez et al., 1998) (Chen and Nie, 1998); and the earliest nanowire nanosenser was first demonstrated IN 2001 (Cui et al., 2001).

With an enhanced perceptive and understanding of brain functioning, improved prognosis as well as therapeutics intended for neurodegenerative disorders for example, Alzheimer's was presented through nanotechnology (Singh et al., 2008). Nanomedicine has the capability to conquer and overcome biological hurdles, to enhance and develop the bioavalability of drugs (Lavan, McGuire et al., 2003), to target disease and infectious sites only as well as for effectual and targeted drug delivery (Babu, et al., 2013)

The current and novel aged medications includes the use of nanoparticles that are composed of biopolymers, substances, metals and ceramic objects as they have the potential to combat against conditions and prerequisites similar to cancer (Farokhzad et al., 2006) as well as to combat against bacteria that are pathogenic for humans and many more characteristics (Morones et al., 2005) (Panacek at al., 2006).

The role of nanomedicine in biomedical research is illustrated in figure 2.2.



Figure 2.2: Nanomedicine Applications in biomedical research (Patra et al., 2018)

2.3 Nanoparticles in Therapeutics

Polymer-based nanoparticles, synthetic as well as natural, present another different approach for therapeutic and beneficial applications due to certain and definite characteristics such as biocompatibility, non-immunogenicity, non-toxicity as well as biodegradability (Crucho and Barros, 2017). **Natural polymer-based** nanoparticles have the potential to overwhelm toxicity hurdles and issues. They also have a potential to bring consequential progression and development in the effectiveness of therapeutical and remedial agents than already available well known treatments (Letchford et al., 2009).

Metallic nanoparticles that are used in medicinal applications are 1–100 nm in size. It is generally made up of cobalt, nickel, iron, gold, silver as well as their respective oxides such as magnetite, maghemite, cobalt ferrite, and chromium dioxide. **Magnetic nanoparticles** have the potential to target a precise location in the body through the help of an exterior magnetic field (Cuenca et al., 2006). The property of super-paramagnetic assist the secure

and stable release of curative agents to the body/cell in addition to appropriate accumulation at the site of target tissue, thus provides a reproducible and secure therapeutic technique (Fan et al., 2011) (Reilly, 2007). Anti-inflammatory action of AgNPs was identified formerly. This property was due to its potential to inhibit interferon- γ , tumor necrosis factor alpha (TNF- α) and to reduce matrix metallo proteinase (MMP) in addition to proinflammatory cytokines (Kirsner et al., 2001) (Tian et al., 2007). Biosynthesized magnetic nanoparticles extensively suppress the intensity of many enzymes present in liver such as alanine transaminase, alkaline phophatase, serum creatinine as well as uric acid in diabetic mice model (Swarnalatha et al., 2012).

Gold nanoparticles (AuNP) are comprehensively utilized metallic nanoparticles, particularly in cancer prognosis and its treatment. AuNPs can be used for targeted delivery of drugs, where the light irradiation has the ability to activate the release of drug (Mura et al., 2016) (Tian et al., 2016). Silver nanoparticles account for approximately more than 23% of all nano based products. It has been extensively used for prognostic as well as in therapeutics (e.g. in healing of wounds, arthritic disease and many more). These nanoparticles have been extensively well-known for their antibacterial, antiviral effects and antifungal effects (Kanav et al., 2016).

2.4 Silver Nanoparticles

AgNPs of diverse shapes and sizes hold distinct catalytic distinctiveness, together with surface plasmon resonance (SPR) as well as strong and vivid toxicity to extensive array of microorganisms. Silver ions along with its various salts have comprehensively been utilized due to their ability of prohibitory consequences against various microorganisms for a long period (Kim et al., 2007). Silver can be used as an antibacterial agent. In wound dressings, topical creams, antiseptic sprays, as well as fabrics, silver can be added (Ahmed et al., 2016). AgNPs has the ability to apply their activity of antibacterial, by performing as antibacterial

complements to antibiotics (Beyth et al., 2015). AgNPs reduce the actions of proinflammatory cytokines, interferon gamma and tumor necrosis factor alpha (Shin et al., 2007).

2.5 Characterization of Silver Nanoparticles

UV-spectrometry is utilized in order to validate the configuration of nanoparticles by observing their SPR. The solution of Silver nitrate is used to make silver nanoparticles (AgNPs). This indicates a highest absorbance of approximately 436 nm (Nithya and Ragunathan, 2009). The morphology (shape) along with particle mass of AgNPs can be identified using transmission electron microscopy, scanning (TEM and SEM) as well as atomic force microscopy (AFM). In addition, dynamic light scattering and X-ray diffraction can be utilized intended for examination of size distribution of particle along with its crystallinity (Nour et al., 2010). Silver nanoparticles can be spherical, rod shaped (Wang et al., 2006), nanowires (Murphy and Jana 2002) and many more shapes.

2.6 Synthesis of Silver Nanoparticles

Different synthetic Silver Nanoparticles methods yield different as well as inconsistent sizes, shapes as well as morphology. They also differ in their stability. Generally, there are three main methods: physical, chemical, and biological/green synthesis given in the figure 2.6:

2.6.1 Physical synthesis

Evaporation/condensation, laser ablation are considered as major substantial approaches for synthesizing and originating nanosilver from the samples like metal. The **evaporation/condensation** approach uses a tube of furnance which is kept under atmospheric pressure to manufacture AgNPs (Jung et al., 2006). **Laser synthesis** approach does not use chemical reagents. It only uses the laser ablation of metals in solution that ultimately leads to pure colloids of nanosilver (Tsuji et al., 2002).



Figure 5.6: Methods for the synthesis of AgNPs (Adnan and Kang 2015)

2.6.2 Chemical synthesis

Chemical reduction is considered as a common process of production of nanosilver. This technique utilizes silver salt, reductants, along with a stabilizer or capping agents. Among all of them, silver nitrate (silver salt) is frequently used for silver nanoparticles synthesis. The reason is that silver nitrate is cost effective as well as it has chemical stability in comparison to various accessible salts of silver (Tien et al., 2008). The reductants contain borohydride (Evanoff and Chumanov, 2005), citrate (Pvatenko et al., 2007), ascorbate (Blanco et al., 2010), and hydrogen gas (Moore, 2006).

2.6.3 Biological synthesis of Silver nanoparticles

In biological strategies, Ag-NPs are synthesized by the use of plants, algae, yeast, fungi, and bacteria as reducing and stabilizing agents in the synthesis (Sintubin et al., 2012).

Shewanella oneidensis which is a metal reducing agent was utilized in order to biologically synthesize Ag-NPs utilizing silver nitrate [Ag (NO₃)] solution as precursor. The resultant

obtained Ag-NPs size was approximately lesser than 15 nm, having consistent dispersion, spherical shape, improved stability, and bulk surface area (Suresh et al., 2012). Stable Ag-NPs which has the size lesser than 20 nm can be made by means of airborne bacteria (*Bacillus* sp.) in which silver nitrate $Ag(NO_3)$ will be used as the precursor (Thirumalai et al., 2010).

In a different study that was illustrated by Venkata Subbaiah and Savithramma, they discovered that Ag-NPs can also be produced from *Cadaba Fruticosa* leaves, in which silver nitrate $Ag(NO_3)$ will be utilized as precursor (Subbaiah and Savithramma, 2013). Some of the plant synthesized silver nanoparticles having antidiabetic potential have been enlisted in the table 2.6.3

Plant	Shape/Size	Plant's Part	Reference
	(nm)		
Cymbopogon	75-138	leaves	Agarwal et al., 2018
citrates			
Alyssum	30	Seeds	Ghasemian Lemraski and
homalocarpum			Valadbeigi 2018
Argyreia nervosa	5-35	leaves	Saratale et al., 2017
Punica granatum	35-60	leaves	Saratale, Shin et al., 2018
Ananas comosus	Spherical	Outer peel of	Gitishree et al., 2019
Muga nanadisia sa	20.60	Stom	ANhozhogon Muumugon et al
Musa paraaisiaca	50-00	Stem	2017
Sonneratia apetala	20-70	Leaf extract	Thatoi et al., 2016
Aloe vera	30	Leaves	Ashraf, Ansari et al., 2016
Heritiera fomes	400	Bark extract	Thatoi et al., 2016
Solanum nigrum	4–25	Leaves	Sengottaiyan et al., 2016
Holoptelea	32-38	Leaves	Kumar et al., 2019
integrifolia			
Withania	14	Leaves	Tripathi et al., 2019
coagulans			

 Table 2.6.3: Green synthesized silver nanoparticles to treat diabetes

2.6.4 Comparison of chemical and biological method

Most common method is chemical method for industrial scale synthesis of Silver nanoparticles. Chemical methods however require stabilizing agent to prevent agglomeration of silver nanoparticles. Although the production rate is fast but chemical method is an intensive and energy consuming method. Contamination of reducing agent during the preparation of silver nanoparticles limits their biomedical application (Krutyakov et al 2008).

Therefore, eco-friendly nanoparticles are needed to be synthesized. Biological synthesis on the other hand is eco-friendly, green process not consuming energy. Therefore, use of organisms and plants for preparation of silver nanoparticles is eco-friendly approach. (Ghorbani et al 2011).

2.7 Role of Silver Nanoparticles for treatment of Diabetes:

Antidiabetic potential of silver nanoparticles (AgNPs) is possibly because of **inhibition of Reactive Oxygen Species (ROS), inflammation** and **enzymes involved in diabetes.** Reactive Oxygen Species (ROS) that includes superoxide radical (.O2), hydrogen peroxide (H2O2) and hydroxyl radical (OH) amplified levels have potential to originate damage in DNA along with prohibition of proteins directly (Chopra at al., 1998). The foremost target substrates to initiate free oxygen radical activity includes polyunsaturated fatty acids, after modification the end product is malondialdehyde (MDA) which is excreted in urine, blood, as well as other body fluids. Consequently it can also serves as a marker of lipid peroxidation along with oxidative stress respectively (Patterson et al., 1998) (Marks et al., 1996).

Abdelazim and Afifi silver (SNPs) tried to find whether silver nanoparticles ameliorate the oxidative stress resulted from diabetes in diabetic rats. A significant increase in the activity and mRNA expression level of superoxide dismutase enzymes (SOD), Catalase (CAT), Glutathione peroxidase (GPx) as well as reductase (*GRd*) was observed. Prabhu et al. in 2017

found the antioxidant potential of AgNPs biologically synthesized silver nanoparticles using Pouteriasapota. The activity of superoxide dismutase enzymes and Catalase was normal in liver tissue homogenates from AgNPs- treated groups (Afifi, M. and A.M. Abdelazim,2015).

Principally two carbohydrate hydrolyzing enzymes (α -amylase and α -glucosidase) are responsible for postprandial hyperglycemia. These two enzymes convert the carbohydrates into monosaccharides, as c consequent of which postprandial hyperglycemia occurs. Therefore inhibitors of α -amylase and α -glucosidase can play an important part in the control of hyperglycemia, because they have the potential to delay digestion of carbohydrate and thus reducing postprandial plasma glucose level(Prabhu et al., 2018) (Hara and Honda 1990). Balan et al. synthesized AgNPs using the aqueous leaf extract of *Lonicera japonica*. These silver nanoparticles inhibit carbohydrate digestive enzymes, thus showing strong anti-diabetic potential (Balan et al., 2016).

AgNPs have been reported in reduction of inflammation that is believed to be the cause of insulin resistance in diabetes Mellitus[58]. Sengottaiyan et al. in 2016 showed that in addition to control blood glucose level and body weight of mice, AgNPs significantly improves the dyslipidemic condition which is clearly observed in the diabetic control (Sengottaiyan et al., 2016).

2.8 Toxicity of Silver Nanoparticles

The toxicity regarding silver nanoparticles (AgNPs) is interrelated with their alteration under biological circumstances as well as environmental media. It also includes their interactions with organic macromolecules, surface oxidation, as well as the release of silver ions (McShan et al., 2014). AgNPs are usually offered as extremely efficient antimicrobial agents which has non-toxic effects to healthy cells of mammals (Stensberg et al., 2011). On the other hand, different in vitro research established the nanosilver-related toxic consequences in hepatocytes of rat as well as neuronal cells (Mahdy et al., 2015), murine stem cells, human lung epithelial cells (Pinzaru et al., 2018).

In a previous oral toxicity investigation of rats, Kim and his co-workers additionally observed that silver nanoparticles can accumulate in blood, liver, lungs, kidneys, stomach, testes, and brain. However AgNPs confirmed no considerable genotoxicity after the silver nanoparticles of approximately 60 nm were orally administered for about 28 days at various concentrations (Kim et al., 2008).

2.9 Anti-oxidant and Pharmacological Properties of Thymus serpyllum

Hussain and his co-workers suggested that *T. serpyllum* essential oils have enhanced radical scavenging activity than *Thymus linearis* essential oil (Hussain et al., 2013). Petrović and his co-workers estimated the antioxidant capability of wild thyme, in which T. serpyllum converts DPPH (1,1-diphenyl-2-picryl-hydrazyl) into its reduced form DPPH-H. In this study, Thymus serpyllum showed even better antioxidant activity than synthetic antioxidants (Petrovic et al., 2014). Mihailović-Stanojević and his co-workers demonstrated the antioxidant potential of wild thyme aqueous extracts in rats' model. In this aqueous extract, phenols and flavonoids were abundantly present, (Mihailovic et al., 2013).

T. Serpyllum Extract	Pharmacological Activity	References
Aqueous	• Anti-hypertensive	(Mihailovic-Stanojevic et al.,
	• Anti-diabetic	2013) (Mushtaq et al., 2016)
Ethanol	Anti-oxidant	(Joshi and Joyal, 2018)
	• Anti-diabetic	(Mushtaq et al., 2016)
Methanol	Anti Cancer	
	• Epigenetic	(Bozkurt et al., 2012)
	modifications	
Ether	Anti-diabetic	(Mushtaq et al., 2016)
Aqueous-Ethanol	Anti tumor	
	• Lipid peroxidation	(Aralbaeva et al., 2017)

Table 2.9: Pharmacological properties of different extracts of T. Serpyllum

Wild thyme in addition to flavonoids and phenol, it also has carboxylic acids along with their derivatives, triterpenes as well as tannins (Wichtl, 2001). Pharmacological properties of different extracts of *Thymus serpyllum* is given in the table 2.7.According to the PDR for Herbal Medicines, the principal constituent of *T. serpyllum* is carvacrol, although it also has borneol, isobutyl acetate, caryophyllene, 1,8-cineole, citral, citronsellal, citronellol, *p*-cymene, geraniol, linalool, α -pinene, γ -terpinene, α -terpineol, terpinyl acetate, and thymol in comparatively elevated concentrations (Thomson, 2004).

CHAPTER 3

Materials and Methods

All the experimentation of the research was done in Atta-ur-Rahman School of Applied Biosciences (ASAB). For the characterization of silver nanoparticles, equipment in School of Chemical and Material Engineering (SCME) at National University of Sciences and Technology (NUST). Details of the methodology are given as follows:

3.1 Docking Analysis3.1.1 Prediction of tertiary structure:

3D structure of GLUT4 was unavailable in Protein Data Bank (PDB) (https://www.rcsb.org/), therefore, it was estimated via online bioinformatics tools. Amino acid sequence of GLUT4 protein was attained from NCBI Database (ncbi.nlm.nih.gov/) (Jenuth, 2000) and GOR4 server (<u>https://npsa-prabi.ibcp.fr/cgi-bin/npsa_automat.pl?page=/NPSA/npsa_gor4.html</u>) (Garnier et al., 1996) was run to build up the secondary structure of this sequence. Software which is TMHMM was adopted to find out the trans membrane domains in GLUT4 protein (http://www.cbs.dtu.dk/services/TMHMM/) (Krogh et al., 2001).

All three tactics of 3D modeling were utilized in order to develop the tertiary structure of GLUT4 i.e. Ab initio, Threading and Homology modeling. FALCON (http://protein.ict.ac.cn/TreeThreader/) and i-TASSER (https://zhanglab.ccmb.med.umich.edu/I-TASSER/) softwares were operated for ab-initio 3D modeling (Wang et al., 2015; Zhang, 2008), whilst MUSTER (https://zhanglab.ccmb.med.umich.edu/MUSTER/) (Wu and Zhang, 2008) and Phyre 2 (http://www.sbg.bio.ic.ac.uk/phyre2/html/help.cgi?id=help/faq) were recruited for threading of based development tertiary IntFOLD structure. (https://www.reading.ac.uk/bioinf/IntFOLD/) (Kelly et al., 2015) and Swiss Model

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(swissmodel.expasy.org/) (Benkert et al., 2010) were employed for homology modeling of GLUT-4 protein in order to construct its 3D tertiary structure.

3.1.2. Validation of Tertiary Structure

Best developed structures by above used software were then compared by exploiting Rampage and SAVES analysis. Rampage examined them on the basis of Ramachandran plot (Lovell et al., 2003) whereas SAVES analyze them on the basis of ERRAT (Colovs and Yeates, 1993), WHATCHECK (Hooft, 1996) and PROCHECK (Laskowski et al., 1996). The best structure was then manipulated for docking scrutiny.

3.1.3 Preparation of 3D structure of Ag

The .SDF file of silver (Ag) was downloaded from Pubchem database (Kim et al., 2015). This file was converted into PDB format by adopting Discovery Studio 2017 (Studio, 2008).

3.1.4. Molecular docking

Molecular docking analysis was executed to observe the binding capability of Ag with GLUT4 protein. PatchDock server (https://bioinfo3d.cs.tau.ac.il/PatchDock/) was utilized to perform the docking (Schneidman et al., 2005). PatchDock is a geometry-based flexible docking method that is accessed to study the biomolecular complexes (Dominguez et al., 2003). RMSD value was set on 4 Å and PDB files of Ag and GLUT4 were uploaded in the designated areas. On the basis of docking scores, docked file were chosen and then uploaded on PDBsum generate to find out the interactions. The result was then visualized through the PyMOL molecular graphic system v.1.3 (Delano, 2002).

After docking of silver with Glut4, *thymus serpyllum* plant mediated silver nanoparticles were then synthesized and characterized.

3.2 Plant Selection and Storage

Thymus serpyllum plant was collected from Rakaposhi Base Camp, Gilgit Baltistan for the

synthesis of silver nanoparticles. The aerial parts of the plant were then dried (figure 3.2).



Figure 3.2: Dried Thymus serpyllum

These parts were then ground into a fine powder, using an automated electric grinder. This powder was then stored at room temperature in a sterile sealed container.

3.3 Preparation of Plant Extract

By the modification of the protocol of Sun et al (Sun, Cai et al., 2014), plant extract was prepared. 10 gram of the plant *Thymus serpyllum* plant powder was soaked in 100 ml deionized water in Erlenmeyer flask at room temperature for few minutes.

This mixture was then heated at 60° C for 15 minutes on a hotplate. The extract was then cooled for about half an hour and then the supernatant was collected. The supernatant was then twice filtered with 0.45 µm pore size filter paper using vacuum filtration assembly. The filtrate obtained was then stored at 4°C as a stock solution. This stock solution can be used within 1 week.

3.4 Synthesis of Silver Nanoparticles using Thymus serpyllum

The stock solution of the plant acts as reducing and capping agent for the precursor. This solution was diluted to 15% (v/v). 850μ l of (10mM) silver nitrate was then added drop wise per second into the 14.25 ml solution taken from 15% diluted extract of *Thymus serpyllum*

under 25°C temperature and continuous magnetic stirring at 300 rpm. This working solution was then kept in rotary orbital shaker in dark for overnight at 700 rpm and 35°C temperature, followed by monitoring using spectrophotometer for SPR band after every 1 hour.

3.5 Purification of Nanoparticles

The reaction mixture was then centrifuged in a refrigerated centrifuge at 15000 rpm for 30 min at 4°C. Supernatant was then discarded. The obtained pellet was re-suspended thrice in deionized water. And it was centrifuged with the same conditions (15000 rpm for 30 min at 4°C) in order to obtain a thicker pellet. This pellet was then air dried, collected and stored in Eppendorf tubes. This dried mass of silver nanoparticles was weighed. The dried mass (silver nanoparticles) was now ready to use for further activities.

3.6 Characterization of Silver Nanoparticles3.6.1 UV Visible Absorption Spectroscopy

In chemical and clinical laboratories, this technique is the most widely used. This technique helps to measure the amount of absorption that occurs in a sample when the beam of light passes through it. In this instrument, the beam of light is split in two parts. One half of the beam is directed towards the cuvette containing the reference solvent (deionized water) and the other half is directed towards the cuvette containing the sample (*thymus serpyllum* mediated silver nanoparticles) to be analyzed. This is a powerful technique in order to analyze the size, stability and the concentration of silver nanoparticles.

Thymus serpyllum mediated silver nanoparticles were first characterized using Spectrophotometer (LABOMED, Inc. U.S.A, Model uvd-2950) in order to observe the specific peak of the silver nanoparticles in the UV visible absorption spectroscopy. The spectra range for absorption was set from 300-600 nm. Spectrophotometer was first blanked by filling both cuvettes with deionized water. After this one cuvette was filled with silver nanoparticle solution and the other with deionized water as reference solvent. Both cuvettes

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were then loaded in the spectrophotometer chamber, where the silver nanoparticles, depending upon the particle size distribution, absorbed the photons of specific wavelength.

3.6.2 X-Ray Diffraction (XRD) Analysis

X-Ray diffraction can be used to study the atomic and molecular structure of silver nanoparticles crystals. XRD produce an intense beam of X-rays which is thrown on the test sample. These X-rays reflect with different intensities and at different angles by the sample. After reflection of X-rays, the data is calculated and a 3D structure is formed by XRD. The angle and the position of an atom in a crystal lattice can be revealed from this 3D structure.

For XRD analysis, the sample was made highly concentrated using the centrifuge. For this, 1ml of the sample was taken in Eppendorf tube and was then centrifuged for about 15 min at 15,000 rpm. Subsequent to 1^{st} centrifugation the supernatant was removed. More sample solution was added in the pellet and then centrifuged again. This process was repeated for about 7-8 time until enlarge thick pellet is formed, which is the important requirement for XRD. Pellet was placed in a glass slide. This slide was then placed under the lamp for 20-25 minutes for drying of the pellet. As a result, a thick layer of silver nanoparticles was deposited on the glass slide for XRD analysis. The glass slide containing the sample was then carefully placed in the XRD machine (STOE, model number theta, Germany). The radiation source used was Copper K alpha, with 40 KV voltage, and 40mA current. The angle used in the process was 20-80 theta (θ).

3.7 Animals for *In vivo* Study

4 weeks old male BALB/c mice (n=20) were taken from animal house of Atta-ur-Rahman School of Applied Biosciences (ASAB), National University of Sciences and Technology (NUST) in which the study was conducted. These male BALB/c mice were kept in cages (5 mice per cage) at constant temperature $(25\pm2^{\circ}C)$ and natural light-dark cycle (12-12 hours) and were initially fed with basic chow diet and were given glucose water.

3.7.1 Ethics Statement

The approval for all the protocols that were carried out during the research was obtained from internal review board (IRB), Atta-ur-Rahman School of Applied Biosciences (ASAB), National University of Sciences and Technology (NUST). All the experiments performed were according to the guidelines provided by the Institute of Laboratory Animal Research, Division on Earth and Life Sciences, National Institute of Health , USA (Guide for the Care and Use of Laboratory Animals: Eighth Edition, 2011).

3.7.1 Antiglycation Assays

3.7.1 BSA-glucose

With the modification of protocol of Zheng et al., 2008, this assay was performed. Briefly, BSA (1200mg) and glucose (1.5mM) were dissolved in PBS (pH 7.4) with 0.1% sodium azide. Test samples containing silver nanoparticles (15, 7.5, 3 and 0.6mg/mL) and aminoguanidine (15, 7.5, 3 and 0.6mg /mL) were added in the above solution. Aminoguanidine was used as positive control. All these samples were then incubated at 37°C for 7 days. After this incubation period, fluorescence intensity was measured at 340nm (excitation wavelength) and 420nm (emission wavelength).

3.7.2 BSA-methyglyoxal

With the modification of protocol of Zheng et al., 2008, this assay was performed. Briefly, BSA (1200mg) and methylglyoxal (4.32mg) were dissolved in PBS (pH 7.4) with 0.1% sodium azide. Test samples containing silver nanoparticles (15, 7.5, 3 and 0.6mg/mL) and aminoguanidine (15, 7.5, 3 and 0.6mg/mL) were added in the above solution. Aminoguanidine was used as positive control. All these samples were then incubated at 37°C for 7 days. After this incubation period, fluorescence intensity was measured at 340nm (excitation wavelength) and 420nm (emission wavelength). To calculate %age inhibition, following formula was used:

Inhibition% =
$$[1-(A_i/A_o)] \times 100$$

 A_i = Fluorescence of BSA and glucose/ methylglyoxal with nanoparticles/aminoguanidine

A_o = Fluorescence of BSA and glucose/ methylglyoxal with phosphate buffer

3.8 Animals for *In vivo* Study

3-4 weeks old male BALB/c mice (n=20) were taken from animal house of Atta-ur-Rahman School of Applied Biosciences (ASAB), National University of Sciences and Technology (NUST) in which the study was conducted. These male BALB/c mice were kept in cages (5 mice per cage) at constant temperature ($25\pm2^{\circ}$ C) and natural light-dark cycle (12-12 hours) and were initially fed with basic chow diet and were given glucose water.

3.8.1 Ethics Statement

The approval for all the protocols that were carried out during the research was obtained from internal review board (IRB), Atta-ur-Rahman School of Applied Biosciences (ASAB), National University of Sciences and Technology (NUST). All the experiments performed were according to the guidelines provided by the Institute of Laboratory Animal Research, Division on Earth and Life Sciences, National Institute of Health , USA (Guide for the Care and Use of Laboratory Animals: Eighth Edition, 2011).

3.8.2 Mice Model Construction to induce T2DM

T2DM mice model was constructed by combination of High Fat Diet (HFD) along with low doses of Streptozotocin. The normal control mice (n=10) were fed with basic chow diet consisting of the following components given in table 3.7.2

Component	Quantity (%)
Crude Fibre	4
Crude Fat	9
Crude Protein	30
Moisture	10.4

Table 3.8.2: Components of Basic Chow Diet

The mice models (n=10) for the purpose of diabetes induction were switched to high fat diet when they were 3 weeks of age. These mice were given intraperitoneal Streptozotocin (100mg/kg) injections prepared in 01.1M citrate buffer (pH 4.5) at 6^{th} and 9^{th} week of age (Day 21 and 42) after overnight fasting. The control animals received citrate buffer injections only. After 2 days of 2^{nd} STZ injection i.e. on 44^{th} day, Fasting blood glucose levels of all mice were checked using EasyGluco Blood Glucose Monitoring System. Mice with Blood Glucose levels greater than 126 mg/dL were considered as diabetic (Dong, Xu et al., 2013).

Group Number	Туре	Administration	No. of mice
1	Control Healthy Mice	Normal Saline	10
2	Diabetic mice	None	10

Table 3.8.3: Mice Grouping

3.8.3.1 Normal Control

Group 1 was assigned as normal group and included 10 BALB/c mice, receiving normal diet and saline water.

3.8.3.2 Experimental Group

Group 2 was assigned as the experimental group for diabetes. The mice in this group were given Streptozotocin injections on day 21 and 42 and fed with high fat diet during the experiment. This group will be used for the comparison with the control to confirm whether diabetes is induced or not.

3.9 Confirmation of type2 diabetes in mice model 3.9.1 Body Weight measurements

Effect on body weight of mice due to high fat diet and Streptozotocin injections was studied for 6 weeks. The average weight of the normal mice was compared to the experimental group.

3.9.2 Fasting Blood Glucose Test

The glucose in the blood of the normal mice as well as Streptozotocin induced diabetic BALB/c mice was monitored on weekly basis. For fasting blood glucose test, the animals were fasted for 8 hours. Then the blood was taken from the tail of mice and measured the glucose level using the glucometer.

3.10 Primer Designing

For the designing of Primers, four different genes were selected namely; GLUT4, INS2, IRS2 and beta actin, where beta actin acts as house-keeping gene. Primers were designed through primer 3 plus software.

3.11 UCSC In-Silico PCR

Validity of selected primers was checked through In-silico PCR of UCSC browser (Rhead et al., 2010). Sequence of Forward and Reverse primers were pasted in designated boxes on the official website of In-silico PCR of UCSC browser (https://genome.ucsc.edu/cgi-bin/hgPcr). Target organism i.e. *mus musculus* (mice), was selected in nominated box, and PCR was run. Primers that showed binding to the target region and showed no off-target binding were chosen for further study. Primer sequences are shown in the table 3.10.

3.12 Primer BLAST

Primer BLAST is a tool to design target-specific primers and to confirm the validity of the primers and in this tool a combination of BLAST and a global alignment algorithm occurs to get a full primer-target alignment. Sequence of Forward as well as Reverse primers were official website pasted in designated boxes on the of Primer BLAST (https://www.ncbi.nlm.nih.gov/tools/primer-blast/index.cgi). Target organism i.e. mus musculus (mice), was selected in nominated box, and Primer BLAST was run.

Primer	Gene	Primer sequence	GC	Melting	Self-	Product
Туре			content	Temp.	complementarity	size (bp)
Forward	GLUT4	TTTGCCCCTCAGT	50%	60.2	3.00	
Primer		CATTCTC				
						243
Reverse	GLUT4	TTCTATTTGCCGTC	50%	60.2	1.00	
Primer		CTCCTG				
Forward	IRS2	CCTTGCTCCTCCA	55%	59.0	2.00	
Primer		CTTCTTC				
						207
Reverse	IRS2	GCCCGAACCTCAA	50%	59.4	2.00	207
Primer		TAACAAC				
Forward	INS2	GACTCCCAGAGGA	60%	59.1	3.00	
Primer		AGAGCAG				
						243
Reverse	INS2	CCAGTAACCACCA	55%	60.0	3.00	213
Primer		GCCCTAA				
Forward	Beta	TGTCCACCTTCCA	61.39	52.38	4.00	
Primer	Actin	GCAGATGT				
						101
Reverse	Beta	AGCTCAGTAACAG	61.53	54.55	4.00	101
Primer	Actin	TCCGCCTAG				

Table 3.11: List of Primers

CHAPTER 4

RESULTS

4.1 Primary Structure of Glut4

Tertiary structure of GLUT-4 was not found in PDB; therefore, it was predicted using bioinformatics tools. Primary sequence of a protein is simply the sequence of its constituent amino acids. Primary structure of Glut-4 protein was downloaded from NCBI database in FASTA format with an accession no of P14142.3 (Figure 4.1)

>sp|P14142.3

MPSGFQQIGSEDGEPPQQRVTGTLVLAVFSAVLGSLQFGYNIGVINAPQKVIEQSYNETWLGR QGPEGPSSIPPGTLTTLWALSVAIFSVGGMISSFLIGIISQWLGRKRAMLVNNVLAVLGGSLMG LANAAASYEMLILGRFLIGAYSGLTSGLVPMYVGEIAPTHLRGALGTLNQLAIVIGILIAQVLG LESLLGTASLWPLLLGLTVLPALLQLVLLPFCPESPRYLYIIQNLEGPARKSLKRLTGWADVSG VLAELKDEKRKLERERPLSLLQLLGSRTHRQPLIIAVVLQLSQQLSGINAVFYYSTSIFETAGV GQPAYATIGAGVVNTVFTLVSVLLVERAGRRTLHLLGLAGMCGCAILMTVALLLLERVPAM SYVSIVAIFGFVAFFEIGPGPIPWFIVAELFSQGPRPAAMAVAGFSNWTSNFIIGMGFQYVAEA MGPYVFLLFAVLLLGFFIFTFLRVPETRGRTFDQISAAFHRTPSLLEQEVKPSTELEYLGPDEN D

Figure 4.1: Primary sequence of GLUT4 in FASTA format, downloaded from NCBI

4.2 Secondary Structure

Primary sequence retrieved from NCBI was used for the prediction of the secondary structure of GLUT4 by implementing GOR4 online tool. Results of GOR4 are shown in Figure 4.2 from where it is clear that this protein has 206 amino acids (40.47%) forming alpha helix, 96(18.86%) and 207 (40.67%) amino acids involved in extended strands and random coil formation respectively. Trans membrane domains were established by TMHMM (Krogh et

al., 2001) software and it showed that there are 12 trans membrane domains with GLUT4

(Figure 4.3).



Figure 4.2: Secondary structure of GLUT4 protein, anticipated by GOR4 software

4.3 Transmembrane Domains (TMHMM)

TMHMM result				
# sp[P146/2]1-509 Length: 509				
# sp[P14672]1-509 Number of bls in TMHs: 262 31291				
# sp[P11672]1-509 Exp number, first 60 Abs: 19.07592				
# sp[P14672]1-509 Total prob of N-in: 0.83293				
# sp/P14672/1-509 POSSIBLE N-term signal sequence				
sp P14672 1-509 TMHMM2.0 inside 1 19				
sp P14672 1-509 TMHMM2.0 TMhelix 20 42				
sp P14672 1-509 TMHMM2.0 outside 43 78				
sp P14672 1-509 TMHMM2.0 TMhelix 79 101				
sp P14672 1-509 TMHMM2.0 inside 102 112				
sp P14672 1-509 TMHMM2.0 TMhelix 113 135				
sp P14672 1-509 TMHMM2.0 outside 136 138				
sp P14672 1-509 TMHMM2.0 TMhelix 139 161				
sp P14672 1-509 TMHMM2.0 inside 162 167				
sp P14672 1-509 TMHMM2.0 TMhelix 168 190				
sp P14672 1-509 TMHMM2.0 outside 191 199				
sp P14672 1-509 TMHMM2.0 TMhelix 200 222				
sp P14672 1-509 TMHMM2.0 inside 223 287				
sp P14672 1-509 TMHMM2.0 TMhelix 288 307				
sp P14672 1-509 TMHMM2.0 outside 308 321				
sp P14672 1-509 TMHMM2.0 TMhelix 322 344				
sp P14672 1-509 TMHMM2.0 inside 345 350				
sp P14672 1-509 TMHMM2.0 TMhelix 351 373				
sp P14672 1-509 TMHMM2.0 outside 374 382				
sp P14672 1-509 TMHMM2.0 TMhelix 383 405				
sp P14672 1-509 TMHMM2.0 inside 406 417				
sp P14672 1-509 TMHMM2.0 TMhelix 418 440				
sp P14672 1-509 TMHMM2.0 outside 441 443				
sp P14672 1-509 TMHMM2.0 TMhelix 444 466				
sp P14672 1-509 TMHMM2.0 inside 467 509				

Figure 4.3: Tran membrane domains in GLUT; established by TMHMM

4.4 Tertiary Structure

Tertiary structure was constructed using homology modeling, Threading and ab-initio approaches. SAVES and RAMPAGE online tools were utilized to choose the best one. Structures that had number of amino acids lower than 509 were excluded at once, no matter how much good they were. Figure 4.4.1 and 4.5.2 shows the 3D model of Ramachandran plot is also determined by using RAMPAGE. Ramachandran Plot of I-TASSER developed model of GLUT4 shows 85.3% amino acids in favored region, 8.5% in allowed and 6.2% in disallowed region.



Figure 4.4.1: 3D structure of GLUT-4, predicted by I-TASSER



Figure 4.4.2: Ramachandran Plot of I-TASSER developed model of GLUT4

In the **Table 4.4** the results of SAVES analysis are given. In this analysis, comparison of different developed structures by using various softwares of 3D modeling was done, the I-TASSER developed model was the best one among all others and it is highlighted.

Table 4.4: Results of SAVES analysis; comparison of different developed structures by

using various softwares of 3D modeling, the best one is highlighted.

Model	(Software)	Amino	Ramachandran	ERRAT	WhatCheck	Procheck
		acid	Plot	(Quality	Error:	Error:
		residues	Residues in fav	Factor	Warning:	Warning:
		in final	region:	A)	Pass	Pass
		structure	In allowed			
			region:			
			In outer region			
Model	Swiss-	451	97.3: 2.4: 0.2			
1	Model					
Model	Swiss-	392	90.5: 7.9: 1.5			
1a	Model					
Model	I-TASSER	509	87.4: 11.6: 1	87.885	6: 11: 30	3: 2: 3
2						
<mark>Model</mark>	<mark>I-TASSER</mark>	<mark>509</mark>	<mark>85.3: 8.5: 6.2</mark>	<mark>92.3868</mark>	<mark>6: 10: 31</mark>	<mark>2: 2: 4</mark>
2a						
Model	IntFOLD	509	80.7: 16.9: 2.4	96.4072	8: 16: 23	6: 1: 2
3						
Model	IntFOLD	509	79.7: 17.1: 3.2	97.6048	8: 17: 23	6: 1: 2
3a						
Model	FALCON	509	79.2: 20: 0.8	92.0408	6: 11: 30	3: 2: 3
4						
Model	FALCON	509	77.6: 22.2: 0.2	88.3576	6: 11: 30	3: 2: 3
4a						
Model	PHYRE2	468	95.9: 2.8: 1.3			
5						
Model	PHYRE2	451	88.9: 8.9: 2.2			
5a						
Model	MUSTER	509	78: 21: 1	86.2705	6: 10: 31	3: 2: 3
6						
Model	MUSTER	509	77: 22.4: 0.6	89.749	6: 11: 30	3: 2: 3
6a						

4.5 Docking Analysis

Molecular Docking was conducted to identify the binding affinity of Ag with the GLUT4 active sites. From docking results, it was speculated that silver is embedded in the active sites of Glut4 (Figure 4.5.1) thus indicating that silver has interactions with Glut4. PDBsum generate showed that asparginine residue of Glut4 has interactions with silver (Figure 4.5.2)



Figure 4.5 Interaction of with the binding pockets of Glut-4 protein

List of protein-ligand interactions				
Hydrogen bonds				
<> A T O M 1>	<> A T O M 2>			
Atom Atom Res Res no. name name no. Chain 2138 ND2 ASN 304 A>	Atom Atom Res Res no. name name no. Chain Distance 3429 Ag LIG 1 _ 2.87			

Figure 4.5.2 List of protein-ligand interactions by PDBsum generate

4.6 Preliminary Confirmation of *Thymus serpyllum* mediated silver NPs

After the addition of 10mM solution of silver nitrate to the aqueous extract of *Thymus serpyllum*, light brown color changes to dark brown gradually with time that visually confirms the formation of silver nanoparticles. After 24 hours of air drying of silver nanoparticles in a petri plate, solid blackish brown powder of silver nanoparticles was obtained as shown in fig 4.6.



Figure 4.6: Purified Silver Naonoparticles

4.7 Characterization of Biogenic Silver Nanoparticles

To analyze the morphology, size and purity of the nanoparticles, the characterization of

Thymus serpyllum mediated silver nanoparticles was performed.

4.7.1 UV Visible Absorption Spectroscopy

UV-Vis spectroscopy shows peak with maximum absorbance at 400nm which indicates the uniform shape of silver nanoparticles as shown in figure 4.7.1. However AgNO3 and extract do not have any peak in this region (250-500nm).



Figure 4.7.1: UV-Vis Spectroscopy of Reaction mixture indicating typical absorbance peak of AgNPs around 400nm.

4.7.2 X-Ray Diffraction (XRD)

X-Ray Diffraction is a technique that identifies the structure imperfections and crystallinity in the sample. The patterns that were obtained by X-Ray Diffraction were analyzed by Jade 8 Software and the JCPDS card number (00-004-0783) was obtained that confirms the crystalline and stale formation of biogenic silver nanoparticles.

The XRD pattern shows that they are present in flat form with their basal planes that are parallel to the substrate. The XRD pattern of the silver nanoparticles is shown in the figure 4.7.2. This pattern indicates six intensive peaks i.e. 28° , 32° , 46° , 55° , 57° and 77° obtained at 20. These are indexed at (111), (200), (220), (311), (222), and (400).



Figure 4.7.2: X-ray Diffraction of Thymus serpyllum mediated silver nanoparticles

4.8 Antiglycation Assays

4.8.1 BSA-glucose

Aminoguanidine gives %age inhibition (80%, 64%, 40% and 12%) at 15mg/mL, 7.5mg/mL,

3mg/mL and 0.6mg/mL. However biogenic silver nanoparticles give 78%, 55%, 28% and

6% at 15mg/mL, 7.5mg/mL, 3mg/mL and 0.6mg/mL as shown in the figure 4.8.1:

4.8.2 BSA-methylglyoxal

Aminoguanidine gives % age inhibition (72%, 62%, 44% and 24%) at 15mg/mL, 7.5mg/mL,

3mg/mL and 0.6mg/mL. However biogenic silver nanoparticles give (68%, 52%, 30% and

16%) at 15mg/mL, 7.5mg/mL, 3mg/mL and 0.6mg/mL as shown in the figure 4.8.2:


Figure 4.8.1: Antiglycation of BSA-Glucose



Figure 4.8.2: Antiglycation of BSA-Methylglyoxal

4.9 Evaluation of Induction of Diabetes Mellitus in BALB/c mice4.9.1 Estimation of Body weight during Diabetic mice model construction

Effect on body weight of mice due to high fat diet and Streptozotocin injections was studied for 6 weeks. The average body weight of normal mice was compared to the diabetic group. The body weight of the diabetic BALB/c mice significantly increased due to High fat diet and Streptozotocin injections. Figure 4.8.1 demonstrates that there is a significant (p<0.05) difference between the body weights of control and diabetic mice groups.



Figure 4.9.1 Measurement of body weight after every 1 week of Induction (P<0.05)

4.9.2 Estimation of Fasting Blood Glucose during Diabetic mice model

construction

Fasting blood glucose levels of the diabetic and control mice were measured every week, in order to confirm that diabetes is successfully induced after 42 days protocol. Figure 4.8.2 depicts that there is a significant (p<0.01) difference in FBG levels of diabetic and control mice groups.





(**P<0.01**)

4.10 Primer Validation

4.10.1 UCSC In-Silico PCR and Primer BLAST

Primers designed through Primer 3 Plus software were validated by Insilico PCR tool of UCSC Browser and Primer BLAST to design target-specific primers for polymerase chain reaction. Results depicted that the chosen primers for β -Actin (House Keeping Gene) Glut4, INS2 and IRS2 binds to the target region well and shows no off-target effects. Results of In-silico PCR are shown in figures 4.9.1.1 - 4.9.1.7.

>chr11:69943361+69943547 187bp TTTGCCCCTCAGTCATTCTC TTCTATTTGCCGTCCTCCTG
TTTGCCCCTCAGTCATTCTCatctggccctaagtattcaagttctgtact
gggtttcacctcctgctctaaaagggaaggtgtccgtcggaaggcagctg
agatctggtcaaacgtccggcctctggtttcaggcacttttaggaaggtg
aagatgaagaagccaagCAGGAGGACGGCAAATAGAA

Figure 4.10.1.1: Insilico PCR results of GLUT4

	Companyo (5' >2')					
	Sequence (S-25)	Length	Tm	GC%	Self complementarity	Self 3' complementarity
ward primer	TTTGCCCCTCAGTCATTCTC	20	57.50	50.00	3.00	0.00
verse primer	TTCTATTTGCCGTCCTCCTG	20	57.31	50.00	2.00	1.00
<u>1_030245725.1</u> PR duct length = 187	EDICTED: Mus musculus solute carrier family '	? (facilitated glucose t	transporter)	, member	4 (Slc2a4), transcript variant X2, r	RNA

Figure 4.10.1.2: Primer-BLAST results of GLUT4

>chr8:11008175+11008381 207bp CCTTGCTCCTCCACTTCTTC GCCCGAACCTCAATAACAAC CCTTGCTCCTCCACTTCTTCtcgctctcgtagtactccagccgcgaggc tgcggcggcgacccccagccgcggatgcctcgtcgccgccgggg gccgcgcaacacgaaaaagcgcttgtggccgtgcttctgcttgcgcaggt agccgcacttgcgcacgctgtggttgttgttgttgttgttgttGTTGTTATTGAGG TTCGGGC

Figure 4.10.1.3: Insilico PCR results of IRS2

etailed primer reports							
Primer pair 1							
	Sequence (5'.>3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity	
Forward primer	CCTTGCTCCTCCACTTCTTC	20	57.89	55.00	2.00	0.00	
Reverse primer	GCCCGAACCTCAATAACAAC	20	57.09	50.00	2.00	0.00	
Products on target te	mplates						
> <u>NM_001081212.2</u> Mu	s musculus insulin receptor substrate 2 (Irs2), mRNA						
product length = 207 Forward primer 1 Template 755	CCTTGCTCCTCCACTTCTTC 20 736						
Reverse primer 1 Template 549	GCCCGAACCTCAATAACAAC 20 568						

Figure 4.10.1.4: Primer-BLAST results of IRS2

Figure 4.10.1.5: Insilico PCR results of INS2

<u>etailed primer reports</u>							
Primer pair 1							
	Sequence (5'->3')	Length	Tm	GC%	Self complementarity	Self 3' complementarity	
Forward primer	GACTCCCAGAGGAAGAGCAG	20	59.18	60.00	3.00	1.00	
Reverse primer	CCAGTAACCACCAGCCCTAA	20	59.01	55.00	3.00	1.00	
Products on target te	mplates						
> <u>NM_001185084.2</u> Mu	s musculus insulin II (Ins2), transcript variant 3, mRN	A					
product length = 243							
Forward primer 1 Template 249	GACTCCCAGAGGAAGAGCAG 20 230						
Reverse primer 1 CC	AGTAACCACCAGCCCTAA 20						

Figure 4.10.1.6: Primer-BLAST results of INS2

>chr5:142903780-142903880 101bp TGTCCACCTTCCAGCAGATGT AGCTCAGTAACAGTCCGCCTAG
TGTCCACCTTCCAGCAGATGTggatcagcaagcaggagtacgatgagtcc
ggcccctccatcgtgcaccgcaagtgcttCTAGGCGGACTGTTACTGAGC
T

Figure 4.10.1.7: Insilico PCR results of Beta Actin

The primers of INS2, IRS2 and Glut 4 that are validated by UCSC In-Silico PCR can further used for in vivo studies to determine the mechanism of action of the biogenic silver nanoparticles to treat diabetes mellitus.

CHAPTER 5

DISCUSSION

Type 2 diabetes mellitus (DM) is a persisting metabolic complication and is distinguished by means of persistent hyperglycemia that is the consequence of either entire or incomplete deficiency of insulin secretion or insulin action. The most commonly used therapeutic agent used for type 2 diabetes mellitus is metformin. However, it has various side effects which include its slow mode of action and gastrointestinal infections. Scientists are therefore trying to overcome these limitations by leading towards the nanobiotechnology. Nanotechnology has provided absolutely innovative concepts and innovative approaches/processes in medicinal drug as well as in dentistry which will facilitate us in making superior, former and sure prognosis and to treat patients with least feasible interventions and without any unfavorable outcomes (Tevatia et al., 2016). The nanomedicine is the appliance of nanotechnology for treatment, prognosis, monitoring, as well as control of disease. Nanomedicine has the potential to overwhelm the consequences of conventional therapies.

In diabetes, nanoparticles are used to monitor blood glucose levels, in bio-imaging and bioimaging as well as for targeted insulin delivery vehicles (Mukhopadhyay and Prosenjit, 2018). Silver nanoparticles have large potential for biomedical applications in diagnostics, imaging and treatment. Silver can be used as an antibacterial agent. In wound dressings, topical creams, antiseptic sprays, as well as fabrics, silver can be added (Ahmed et al., 2016). AgNPs reduce the actions of proinflammatory cytokines, interferon gamma and tumor necrosis factor alpha (Shin et al., 2007). However, toxic effects of silver nanoparticles should not be ignored.

It is important to evaluate the dimension of nanoparticles so that they can be used in therapeutics and biomedical research as in an in-vivo studies by Zang et al, have confirmed that nanoparticles with size greater than 80 nm gets entrapped in liver and can exhibit long term cytotoxicity. However, small nanoparticles with size less than 10 nm can excreted via urine. (Zang et al 2014).

In the recent research, the synthesis of nanoparticles by plants have gained much importance because of the presence of various phytochemicals in plants that act as capping and reducing agents (Taghikhani et al., 2014). *Thymus serpyllum* is a significant origin of therapeutic substances that have antioxidant, anti-diabetic, antimicrobial, antitumor potential, as well as it has utility in pharmaceutical, food, and cosmetic industries (Jaric et al., 2015). Mihailović-Stanojević and his co-workers illustrated the antioxidant potential of wild thyme aqueous extracts in rats' model. Phenols and flavonoids were abundantly present in the extract (Mihailovic et al., 2013).

The aims of this study were to dock silver with GLUT4, to synthesize green, cost effective silver nanoparticles (AgNPs) from the extract of *Thymus serpyllum*. This synthesis method was also optimized under different physiochemical parameters such as temperature and molar concentration ratios to obtain narrow size distribution of AgNPs. XRD was used to characterize these silver nanoparticles. To induce diabetes in BALB/c mice, streptozotocin (STZ) injections were given along with high fat diet. Primers for Glut4, INS2, and IRS2 were designed by Primer 3 Plus and then validated by UCSC In-Silico PCR.

Molecular Docking was conducted to identify the binding affinity of Ag with the GLUT4 active sites. Depending upon the docking pose, it was speculated that the Ag demonstrated strong interactions with the active site of GLUT4 and their nearby residues.

By the modification of the protocol of Sun et al (Sun, Cai et al., 2014), silver nanoparticles were synthesized from *Thymus serpyllum* plant extract. The protocol was optimized by varying the concentration of plant extract, silver nitrate and temperature. The bioactive

compounds such as flavonoids, phenols etc present in *Thymus serpyllum* (Mihailovic et al., 2013) act as stabilizing agent during the synthesis of silver nanoparticles.

After overnight incubation, the light yellow shade of the reaction mixture was altered to blackish brown which indicated the preliminary confirmation of synthesis of *Thymus serpyllum* mediated silver nanoparticles. The optimal conditions for *Thymus serpyllum* mediated silver nanoparticles were 35°C with overnight incubation for 14.25ml diluted *Thymus serpyllum* extract and 10mM silver nitrate.

These biogenic silver nanoparticles were initially characterized by UV-VIS spectroscopy, showing maximum absorbance at the characteristic wavelength of 400nm. The shape of these silver nanoparticles was found spherical. X-ray Diffraction pattern of these biogenic silver nanoparticles showed six intensive peaks. Similar peaks are also cited in the literature for the silver nanoparticles (Hu, Wang et al., 2008).

The second objective of the study was the induction of diabetes mellitus in BALB/c male mice. In the study by Magalhães et al, they have also successfully induced diabetes mellitus type 2 with streptozotocin injections combined with High fat diet (Magalhães et al., 2019). Male BALB/c mice were divided into a control group (normal feed with saline water), and experimental group (High fat diet and Streptozotocin injections). Intraperitoneal streptozotocin injections were given to experimental male BALB/c mice on day 21 and 42. Fasting blood glucose and weight of the control and experimental mice were monitored every week. As a result of Streptozotocin injections and high fat diet, the fasting blood glucose level and weight of male BALB/c mice was greater than the control, which indicated the induction of type 2 diabetes mellitus in BALB/c mice.

Next objective was primer designing and their validation. The genes related to obesity and reactive oxygen species i.e. Glut4, INS2 and IRS2 were selected for primer designing. β -actin was used as house-keeping gene. Primer 3 plus was used to design primers. The primers with

good GC content and melting point were selected. These were further validated by UCSC In-Silico PCR. UCSC In-Silico PCR gives the primers that showed binding to the target region and showed no off-target binding and hence selected for further in vivo analysis.

Thus, silver nanoparticles having anti-diabetic potential can be synthesized from *Thymus serpyllum* extract under optimized conditions and type 2 diabetes mellitus can be induced by using streptozotocin injections and high fat diet.

CHAPTER 6

CONCLUSION AND FUTURE PROSPECTS

This study addresses the biosynthesis of anti-diabetic silver nanoparticles using *Thymus serpyllum* plant extract in order to minimize the side effects of synthetic drugs used for the treatment of diabetes. The aqueous extract of *Thymus serpyllum* is rich in phenols, flavonoids and many other bioactive compounds that act as capping and reducing agents. These *Thymus serpyllum* mediated silver nanoparticles were characterized by UV-Visible Absorption Spectroscopy showing maximum absorbance at the characteristic wavelength of 400nm and X-ray Diffraction.

Intraperitoneal streptozotocin injections and continuous feeding of high fat diet to experimental male BALB/c mice induces type 2 diabetes mellitus in male mice. Our results of UCSC In-Silico PCR of Glut4, INS2 and IRS2 genes suggest that these primers show binding to the target region in mice genome and showed no off-target binding and therefore can be used for further in-vivo study.

There is a need of investigation of INS2, Glut4, IRS2 and other genes involved in diabetes to explain the exact mechanism of action of biogenic silver nanoparticles. In future, the testing of anti-diabetic potential of these biogenic silver nanoparticles in diabetic cell line is also needed. Blood Brain Barrier permeability of *Thymus serpyllum* mediated silver nanoparticles for their toxicological assessment.

CHAPTER 7

<u>REFERENCES</u>

- A. K. Suresh, D. A. Pelletier, W. Wang et al., (2010) "Silver nanocrystallites: biofabrication using shewanella oneidensis, and an evaluation of their comparative toxicity on gramnegative and gram-positive bacteria," *Environmental Science and Technology*, vol. 44, no. 13, pp. 5210–5215.
- A. Panacek, L. Kvitek, R. Prucek, M. Kolar, R. Vecerova, N. Pizurova, V. K. Sharma, T. Nevecna, R. Zboril, (2006). J. Phys. Chem. B, 110, 16248.
- Aamir, A. H., Ul-Haq, Z., Mahar, S. A., Qureshi, F. M., Ahmad, I., Jawa, A., Sheikh, A., Raza, A., Fazid, S., Jadoon, Z., Ishtiaq, O., Safdar, N., Afridi, H., & Heald, A. H. (2019). Diabetes Prevalence Survey of Pakistan (DPS-PAK): prevalence of type 2 diabetes mellitus and prediabetes using HbA1c: a population-based survey from Pakistan. *BMJ open*, 9(2), e025300.
- Abdul-Ghani MA, Matsuda M, Jani R, et al. (2008). The relationship between fasting hyperglycemia and insulin secretion in subjects with normal or impaired glucose tolerance. *Am J Physiol Endocrinol Metab.* ;295:E401–E406.
- Afifi, M. and A.M. Abdelazim, (2015). Ameliorative effect of zinc oxide and silver nanoparticles on antioxidant system in the brain of diabetic rats. *Asian Pacific Journal* of Tropical Biomedicine. 5(10): p. 874-877
- Akash MS, Shen Q, Rehman K, Chen S. (2015). Interleukin-1 receptor antagonist: a new therapy for type 2 diabetes mellitus. *J Pharm Sci.* ;101:1647–58
- American Diabetes Association (2010). Diagnosis and classification of diabetes mellitus. *Diabetes care*, *33 Suppl 1*(Suppl 1), S62–S69.

American Diabetes Association. Diagnosis and classification of diabetes mellitus. (2014) Diabetes Care. ;37 Suppl 1:S81–S90.

American Diabetes Association. Standards of medical care in diabetes - 2017

- Anbazhagan, P., K. Murugan, A. Jaganathan, V. Sujitha, C. M. Samidoss, S. Jayashanthani,
 P. Amuthavalli, A. Higuchi, S. Kumar and H. Wei (2017). "Mosquitocidal, antimalarial and antidiabetic potential of *Musa paradisiaca* synthesized silver nanoparticles: in vivo and in vitro approaches". *Journal of Cluster Science* 28(1): 91=107
- Aralbaeva, A. N., Mamataeva, A. T., Zhaparkulova, N. I., Utegalieva, R. S., Khanin, M., Danilenko, M., & Murzakhmetova, M. K. (2017). A composition of medicinal plants with an enhanced ability to suppress microsomal lipid peroxidation and a protective activity against carbon tetrachloride-induced hepatotoxicity. *Biomedicine & Pharmacotherapy*, 96, 1283-1291.
- Arthur C. Chapter 78: Insulin, Glucagon, and Diabetes Mellitus. Textbook of Medical Physiology (11th ed.). Philadelphia: Elsevier Saunders 2006. P 963–P968
- Asadi SY, Parsaei P, Karimi M, Ezzati S, Zamiri A, Mohammadizadeh F, et al. (2013). Effect of green tea (*Camellia sinensis*) extract on healing process of surgical wounds in rat. *Int J Surg.* ;11:332–7
- Ashraf, J. M., M. A. Ansari, H. M. Khan, M. A. Alzohairy and I. Choi (2016). "Green synthesis of silver nanoparticles and characterization of their inhibitory effects on AGEs formation using biophysical techniques." *Scientific reports* 6: 20414
- Asiyanbola, B. and W. Soboyejo (2008). "For the surgeon: an introduction to nanotechnology." Journal of education and health promotion.

- Ay A, Alkanli N, Sipahi T, Gulyasar T, et al. (2018). Investigation of the relationship between MTHFR, IRS and CALCA gene polymorphisms and development of diabetic nephropathy in patients with type 2 diabetes mellitus. *Biotechnology & Biotechnological Equipment* 1-9.
- Babu, A., A. K. Templeton, A. Munshi and R. Ramesh (2013). "Nanoparticle based drug delivery for therapy of lung cancer: progress and challenges." *Journal of Nanomaterials* : 14
- Bajaj, S., & Khan, A. (2012). Antioxidants and diabetes. Indian journal of endocrinology and metabolism, 16(Suppl 2), S267–S271.
- Balan, K., et al., (2016). Antidiabetic activity of silver nanoparticles from green synthesis using Lonicera japonica leaf extract. *Rsc Advances*. 6(46): p. 40162-40168
- Beladi-Mousavi SS, Bashardoust B, Nasri H, Ahmadi A, Hajian S, Torkamaneh S. (2014). Association of serum magnesium with Vitamin D level in normal and renal disease patients. *J Parathyr Dis.* ;2:45–6.
- Belkina AC, Denis GV. (2010). Obesity genes and insulin resistance. *Curr Opin Endocrinol Diabetes Obes.* ;17(5):472–477
- Benítez G., González-Tejero M. R., Molero-Mesa J. (2010). Pharmaceutical ethnobotany in the western part of Granada province (southern Spain): ethnopharmacological synthesis. *Journal of Ethnopharmacology*. ;129(1):87–105.
- Blanco-Andujar C, Tung LD, Thanh NTK. (2010). Synthesis of nanoparticles for biomedical applications. *Annual Reports Section "A" (Inorganic Chemistry)*.;106:553–568.

- Bozkurt, E., Atmaca, H., Kisim, A., Uzunoglu, S., Uslu, R., & Karaca, B. (2012). Effects of Thymus serpyllum extract on cell proliferation, apoptosis and epigenetic events in human breast cancer cells. *Nutrition and cancer*, 64(8), 1245-1250.
- Chaturvedi N. (2007). The burden of diabetes and its complications: Trends and implications for intervention. *Diabetes Res Clin Pract.* ;76(3):S3–S12
- Chen L, Magliano DJ, Zimmet PZ (2011). The worldwide epidemiology of type 2 diabetes mellitus-present and future perspectives. *Nat Rev Endocrinol.* ;8(4):228–236.
- Cho NH, Shaw JE, Karuranga S, et al. (2018). IDF Diabetes Atlas: Global estimates of diabetes prevalence for 2017 and projections for 2045. *Diabetes Res Clin Pract* ;138:271–81.
- Corbett, J., P. McKeown, G. Peggs and R. Whatmore (2000). "Nanotechnology: international developments and emerging products." *CIPS Annals* 49(2):523-545
- Crucho, C.I.C. (2017). Barros, M.T. Polymeric nanoparticles: A study on the preparation variables and characterization methods. *Mater. Sci Eng. C Mater. Biol. Appl.* 80, 771– 784.
- Cuenca, A.G.; Jiang, H.; Hochwald, S.N.; Delano, M.; Cance, W.G.; Grobmyer, S.R. (2006). Emerging implications of nanotechnology on cancer diagnostics and therapeutics. *Cancer.* 107, 459–466.
- D. Tripathi, A. Modi, G. Narayan, S.P. Rai (2019). Green and cost effective synthesis of silver nanoparticles from endangered medicinal plant *Withania coagulans* and their potential biomedical properties. *Mater. Sci. Eng. C.*, 100: 152-164
- Das, G., Patra, J.K., Debnath, T., Ansari, A., Shin, S.-H. (2019). Investigation of antioxidant, antibacterial, antidiabetic, and cytotoxicity potential of silver

nanoparticles synthesized using the outer peel extract of *Ananas comosus* (L.). *PLoS ONE* 14(8): e0220950

- Dedon PC, Tannenbaum SR. (2004). Reactive nitrogen species in the chemical biology of inflammation. *Arch Biochem Biophys* ;423:12–22
- DeFronzo RA. (1988). The triumvirate : B-cell, muscle, liver. A collusion responsible for NIDDM. *Diabetes*; 37: 667-87.
- DeFronzo, R. A., Ferrannini, E., Groop, L., Henry, R. R., Herman, W. H., Holst, J. J., ... & Simonson, D. C. (2015). Type 2 diabetes mellitus. *Nature reviews Disease primers*, 1, 15019.
- Desiderio A, Spinelli R, Ciccarelli M, Nigro C, et al. (2016) Epigenetics: spotlight on type 2 diabetes and obesity. *Journal of Endocrinological Investigation*. 39: 1095-1103.
- Dhingra R, Sullivan L, Jacques PF. et al. (2007). Soft drink consumption and risk of developing cardio-metabolic risk factors and the metabolic syndrome in middle-aged adults in the community. *Circulation*; 116:480–488.
- Donadon V, Balbi M, Casarin P. et al. (2008). Association between hepatocellular carcinoma and type 2 diabetes mellitus in Italy: potential role of insulin. *World J Gastroenterol*; 14(37):5695–5700
- Ehses JA, Perren A, Eppler E et al. (2007). Increased number of islet-associated macrophages in type 2 diabetes. *Diabetes*; 56:2356–70
- Erisonn J, Taimola S, Eriksson K, Parviainen S, Peltonen J, Kujula U. (1997) Resistance training in the treatment of non-insulin dependent diabetes mellitus. *Int J Sports Med.* 18:242–246

- Etchegaray JP and Mostoslavsky R (2016) Interplay between Metabolism and Epigenetics: A Nuclear Adaptation to Environmental Changes. *Molecular Cell*; 62: 695-711.
- Evanoff DD Jr, Chumanov G. (2005). Synthesis and optical properties of silver nanoparticles and arrays. *Chemphyschem*;6(7):1221–1231.
- Fan, C.; Gao, W.; Chen, Z.; Fan, H.; Li, M.; Deng, F.; Chen, Z. (2011). Tumor selectivity of stealth multi-functionalized superparamagnetic iron oxide nanoparticles. *Int J. Pharm*, 404, 180–190
- Feynman RP. (1960). There's plenty of room at the bottom. Eng Sci Feb; 23:22-36.
- Franz MJ, Horton ES, Bantle JP, et al. (1994). Nutrition principles in the management of diabetes and related complications. *Diabetes Care*. 17:490–518
- Galtier F. (2010). Definition, epidemiology, risk factors. Diabetes Metab.; 36:628-651
- Garhwal, S. 2010. Medicinal plants as a source of antioxidants. *Research Journal of Medicinal plants and By product;* 7(1):1–8
- GBD 2017 Disease and Injury Incidence and Prevalence Collaborators. Global, regional, and national incidence, prevalence, and years lived with disability for 354 diseases and injuries for 195 countries and territories, 1990–2017: a systematic analysis for the Global Burden of Disease Study 2017. *Lancet Lond Engl* 2018; 392(10159):1789–858
- Genetic basis of type 1 and type2 diabetes, obesity, and their complications (2011). Advances and emerging opportunities in diabetes research: a Strategic Planning report of the DMICC
- Ghamarian A, Abdollahi M, Su X, Amiri A, Ahadi A, Nowrouzi A. (2012).Effect of chicory seed extract on glucose tolerance test (GTT) and metabolic profile in early and late stage diabetic rats. *Daru* :20–56.

- Ghasemian Lemraski, E. and T. Valadbeigi (2018). "Evaluation of in vitro Antimicrobial, Antidiabetic and Antioxidant Potential of Alyssum homalocarpum and green synthesis of silver nanoparticles." *Journal of Medicinal Plants and By products*; 7(1):1-8
- Sarah W. Gojka R. Anders G., Richard S. and Hilary, K. (2004). Global Prevalence of Diabetes Estimates for the year 2000 and projections for 2030. *Diabetes Care*; 27(5): 1047-1053.
- Happy Agarwal, S. Venkat Kumar, S. Rajeshkumar (2018). Antidiabetic effect of silver nanoparticles synthesized using lemon grass (*Cymbopogon citratus*) through onventional heating and microwave irradiation approach. *Journal of Microbiology, Biotechnology and Food sciences;* 7(4)
- Hara, Y. and M. Honda. (1990). *The* inhibition of α-amylase by tea polyphenols. *Agricultural and Biological Chemistry*, **54**(8): p. 1939-1945.
- Harley M. R., Atkins S., Budantsev L. A., et al. Labiatae. In: Kubitzki K., Kadereit J. W., editors. (2004). *The Families and Genera of Vascular Plants*. Vol. 6. Berlin, Germany: Springer; pp. 167–275.
- Heidarian E, Rafieian-Kopaei M. (2013). Protective effect of artichoke (*Cynara scolymus*) leaf extract against lead toxicity in rat. *Pharm Biol.*; 51:1104–9.
- Helmrich SP, Ragland DR, Paffenberger RS. (1994). Prevention of non-insulin diabetes mellitus with physical activity. *Med Sci Sports Exerc*. 26:824–850
- Henry RR, Gumbiner B. (1991). Benefits and limitations of very low calorie diet therapy in obese NIDDM. *Diabetes Care*. 14:802–823
- Holleman F, Hoekstra JBL. (1997). Insulin Lispro. N Engl J Med. 337:176-183

- Hu FB, Manson JE, Stampfer MJ. et al. (2001). Lifestyle, and the risk of type 2 diabetes mellitus in women. *N Engl J Med.*;345(11):790–797
- Hulbert AJ, Turner N, Storlien LH, Else PL. (2005). Dietary fats and membrane function: Implications for metabolism and disease. *Biol Rev Camb Philos Soc.* ;80:155–69
- Hung HY, Qian K, Morris-Natschke SL, Hsu CS, Lee KH. (2012). Recent discovery of plantderived anti-diabetic natural products. *Nat Prod Rep.* ;29:580–606
- Hussain A. I., Anwar F., Chatha S. A. S., et al. (2013). Chemical composition and bioactivity studies of the essential oils from two *Thymus* species from the Pakistani flora. *LWT*— *Food Science and Technology*.;50(1):185–192.
- Ismail-Beigi F, Craven T, Banerji MA, Basile J, et al. (2010) Effect of intensive treatment of hyperglycaemia on microvascular outcomes in type 2 diabetes: an analysis of the ACCORD randomised trial. *Lancet* 376: 419-430.
- Israili, Zafar H PhD (2011). Advances in the Treatment of Type 2 Diabetes Mellitus, American Journal of Therapeutics 18 (2) :p117–152
- J. R. Morones, J. L. Elechiguerra, A. Camacho, K. Holt, J. B. Kouri, J. T. Ramirez, M. J. Yacaman, (2005). *Nanotechnology* 16, 2346.
- J. S. Kim, E. Kuk, K. N. Yu et al., (2007). "Antimicrobial effects of silver nanoparticles," *Nanomedicine: Nanotechnology, Biology and Medicine*, vol. 3, no. 1, pp. 95–101.

Jain KK (2008) The handbook of nanomedicine. Humana Press, Totowa, NJ, USA.

Jakus V. (2000). The role of free radicals, oxidative stress and antioxidant systems in diabetic vascular disease. *Bratisl Lek Listy*.;101:541–51

- Jarić, S., Mitrović, M., & Pavlović, P. (2015). Review of Ethnobotanical, Phytochemical, and Pharmacological Study of Thymus serpyllum L. *Evidence-based complementary and alternative medicine : eCAM*, 2015, 101978.
- Joshi, T., & Juyal, V. (2018). Evaluation of hydroxyl radical scvenging activity of ethanolic extract of Thymus serpyllum. *Int. J. Pharm. Sci. Rev. Res, 9*, 1625-1627.
- Jung JH, Oh HC, Noh HS, Ji JH, Kim SS. (2006). Metal nanoparticle generation using a small ceramic heater with a local heating area. *J Aerosol Sci.*;37(12):1662–1670.
- K. M. M. Abou El-Nour, A. A. Eftaiha, A. Al-Warthan, and R. A. A. Ammar, (2010).
 "Synthesis and applications of silver nanoparticles," *Arabian Journal of Chemistry*, vol. 3, no. 3, pp. 135–140.
- K. P. Venkata Subbaiah and N. Savithramma, (2013). "Synthesis of silver nanoparticles and antimicrobial activity from *Cadaba fruticosa*—an important ethnomedicinal plant to treat Vitiligo of kurnool district, andhra pradesh," *Indo American Journal of Pharmaceutical Research*, vol. 3, no. 12, pp. 1285–1292.
- Kaehler T. (1994). Nanotechnology, Basic concepts and Definitions. *Clinical Chem*; 40(9):1797-99
- Kaira, Laxman & singh, dr. (2012). nanotechnology- the new era of technology.. nitte university *journal of health sciences*. 2. 88-92.
- Kim YS, Kim JS, Cho HS, et al. (2008). Twenty-eight-day oral toxicity, genotoxicity, and gender-related tissue distribution of silver nanoparticles in Sprague-Dawley rats. *Inhal Toxicol.*; 20(6):575–583.
- Kirsner R, Orsted H, Wright B. (2001). Matrix metalloproteinase in normal and impaired wound healing:a potential role of monocrystalline silver. *Wounds*; 13:5-10.

- Krukemeyer MG1*, Krenn V2, Huebner F1, Wagner W1 and Resch R3 (2015). History and Possible Uses of Nanomedicine Based on Nanoparticles and Nanotechnological Progress J Nanomed Nanotechnol; 6:6
- Krutyakov, Y. A., Kudrinskiy, A. A., Olenin, A. Y. and Lisichkin, G. V. (2008). Synthesis and properties of silver nanoparticles: advances and prospects. *Russian Chemical Reviews.*, 77: 233257
- Kumar V., Singh S., Srivastava B., Bhadouria R., Singh R. (2019) Green synthesis of silver nanoparticles using leaf extract of Holoptelea integrifolia and preliminary investigation of its antioxidant, anti-inflammatory, antidiabetic and antibacterial activities. *Journal of Environmental Chemical Engineering*, 7 (3)
- L. Sintubin, W. Verstraete, and N. Boon, (2012). "Biologically produced nanosilver: current state and future perspectives," *Biotechnology and Bioengineering*, vol. 109, no. 10, pp. 2422–2436.
- Lai LL, Wan Yusoff WNI, Vethakkan SR, Nik Mustapha NR, Mahadeva S, Chan WK. (2019). Screening for non-alcoholic fatty liver disease in patients with type 2 diabetes mellitus using transient elastography. J. Gastroenterol. Hepatol; 34(8):1396-1403
- Burge MR, Schade DS. 1997 Insulins. Endocrinol Metab Clin North Am. 26:575-598
- Larsson SC, Andersson SO, Johansson JE. et al. (2008). Diabetes mellitus, body size and bladder cancer risk in a prospective study of Swedish men. *Eur J Cancer*; 44:2655–2660
- Larsson SC, Wolk A. (2011). Diabetes mellitus and incidence of kidney cancer: a metaanalysis of cohort studies. *Diabetologia*; 54:1013–1018

- Lavan, D. A., T. McGuire and R. Langer (2003)."Small-scale systems for in vivo drug delivery." Nature biotechnology 21(10): 1184
- Lee, P.G. and J.B. Halter (2017). The pathophysiology of hyperglycemia in older adults clinical considerations. *Diabetes Care*: 40(4) 444–452
- Letchford, K.; Liggins, R.; Wasan, K.M.; Burt, H. (2009). In vitro human plasma distribution of nanoparticulate paclitaxel is dependent on the physicochemical properties of poly(ethylene glycol)-block-poly(caprolactone) nanoparticles. *Eur. J. Pharm. Biopharm.*, 71, 196–206.
- Lushchak, O., Zayachkivska, A., & Vaiserman, A. (2018). Metallic Nanoantioxidants as Potential Therapeutics for Type 2 Diabetes: A Hypothetical Background and Translational Perspectives. *Oxidative medicine and cellular longevity*, 3407375.
- Magalhães DA, Kume WT, Correia FS, et al. (2019). High-fat diet and streptozotocin in the induction of type 2 diabetes mellitus: a new proposal. *An Acad Bras Cienc.*; 91(1):e20180314. Published 2019 Mar 21.
- Manu MS, Rachana KS, Advirao GM (2017) Altered expression of *IRS2* and GRB2 in demyelination of peripheral neurons: Implications in diabetic neuropathy. *Neuropeptides* 62: 71-79.
- Martin IK, Wahren J. (1993). Glucose metabolism during physical exercise in patients with non-insulin dependent (type 2) diabetes. *Adv Exp Med Biol*. 334:221–233
- Martinez LC, Sherling D, Holley A. (2019). The Screening and Prevention of Diabetes Mellitus. *Prim. Care*; 46(1):41-52
- Mati E., de Boer H. (2011). Ethnobotany and trade of medicinal plants in the Qaysari Market, Kurdish Autonomous Region, Iraq. *Journal of Ethnopharmacology*; 133(2):490–510.

- Matsuda M, DeFronzo RA. (1999). Insulin sensitivity indices obtained from oral glucose tolerance testing: comparison with the euglycemic insulin clamp. *Diabetes Care*; 22:1462–1470.
- Matsui, T., et al., (2007). α-Glucosidase inhibitory profile of catechins and theaflavins. Journal of Agricultural and Food Chemistry, **55**(1): p. 99-105.
- McShan D., Ray P.C., Yu H. (2014). Molecular toxicity mechanism of nanosilver. J. Food Drug Anal. ; 22:116–127.
- Menković N., Šavikin K., Tasić S., et al. (2011). Ethnobotanical study on traditional uses of wild medicinal plants in Prokletije Mountains Montenegro. *Journal of Ethnopharmacology*; 133(1):97–107.
- Mihailovic-Stanojevic N., Belščak-Cvitanović A., Grujić-Milanović J., et al. (2013). Antioxidant and antihypertensive activity of extract from *Thymus serpyllum* L. in experimental hypertension. *Plant Foods for Human Nutrition*; 68(3):235–240.
- Mohamed El Mahdy M., Salah T., Sayed Aly H., Mohammed F., Shaalan M. (2015). Evaluation of hepatotoxic and genotoxic potential of silver nanoparticles in albino rats. *Exp. Toxicol. Pathol.*; 67:21–29.
- Moore K. (2006). A new silver dressing for wounds with delayed healing. *Wounds UK*; 2(2):70–78.
- Mura, S.; Nicolas, J.; Couvreur, P. (2013). Stimuli-responsive nanocarriers for drug delivery. *Nat. Mater.*, 12, 991–1003.
- Murphy, C. J. and N.R. Jana (2002). "Controlling the aspect ratio of inorganic nanorods and nanowires." *Advanced Materials*; 14(1):80-82

- Mushtaq, M. N., Bashir, S., Ullah, I., Karim, S., Rashid, M., & Hayat Malik, M. N. (2016).
 Comparative hypoglycemic activity of different fractions of Thymus serpyllum L. in alloxan induced diabetic rabbits. *Pakistan journal of pharmaceutical sciences*, 29(5).
- N. Beyth, Y. Houri-Haddad, A. Domb, W. Khan, and R. Hazan, (2015). "Alternative antimicrobial approach: nano-antimicrobial materials," *Evidence-Based Complementary and Alternative Medicine*.
- N. Taniguchi (1974). On the Basic Concept of 'Nano-Technology. Proc. Intl. Conf. Prod. London, Part II British Society of Precision Engineering.

Nanotechnology: A Gentle Introduction to the Next Big Idea, by MA Ratner, D Ratner. 2002.

- Nasri H, Behradmanesh S, Maghsoudi AR, Ahmadi A, Nasri P, Rafieian-Kopaei M. (2014). Efficacy of supplementary Vitamin D on improvement of glycemic parameters in patients with type 2 diabetes mellitus; a randomized double blind clinical trial. J Renal Inj Prev. ; 3:31–4
- Nasri H, Rafieian-Kopaei M. (2013). Tubular kidney protection by antioxidants. *Iran J Public Health*; 42:1194–6
- Nasri, H., Shirzad, H., Baradaran, A., & Rafieian-Kopaei, M. (2015). Antioxidant plants and diabetes mellitus. *Journal of research in medical sciences : the official journal of Isfahan University of Medical Sciences*, 20(5), 491–502.
- Patra, J.K., Das, G., Fraceto, L.F. *et al.* Nano based drug delivery systems: recent developments and future prospects. *J Nanobiotechnol* **16**, 71 (2018).
- Peng, X.F., Zheng, Z.P., Cheng, K.W., Shan, F., Ren, G.X., Chen, M.F. (2008). Inhibitory effect of mung bean extract and its constituents vitexin and isovitexin on the formation of advanced Glycation end products. *Food Chemistry*, 106, pp. 475-481

- Petersen KF. (2002). Pathogenesis of skeletal muscle insulin resistance in type 2 diabetes mellitus. *Am J Cardiol*, 90 (5A) 11G 18G
- Petrović S. S., Ristić M. S., Petr N. V., Lazić M. L., Francišković M., Petrović S. D. (2014). Chemical composition and antioxidative activity of essential oil of *Thymus serpyllum* L. *Hemijska Industrija*; 68(3):389–397.
- Pierce M, Keen H, Bradley C (1995) Risk of Diabetes in Offspring of Parents with Non- insulin- dependent Diabetes. *Diabetic Medicine*; 12: 6-13.
- Pieroni A., Rexhepi B., Nedelcheva A., et al. (2013). One century later: the folk botanical knowledge of the last remaining Albanians of the upper Reka Valley, Mount Korab, Western Macedonia. *Journal of Ethnobiology and Ethnomedicine*; 9(1, article 22)
- Pinzaru I., Coricovac D., Dehelean C., Moaca E.A., Mioc M., Baderca F., Sizemore I., Brittle S., Marti D., Calina C.D., et al. (2018). Stable peg-coated silver nanoparticles—A comprehensive toxicological profile. *Food Chem. Toxicol.*; 111:546–556.
- Prabhu, S., et al., (2018). Evaluation of antidiabetic activity of biologically synthesized silver nanoparticles using Pouteria sapota in streptozotocin-induced diabetic rats. *Journal of Diabetes*, **10**(1): p. 28-42.
- Pradhan AD, Manson JE, Rifai N et al. (2001). C-reactive protein, interleukin 6, and risk of developing type 2 diabetes mellitus. *JAMA*; 286:327–34.
- Pyatenko A, Yamaguchi M, Suzuki M. (2007). Synthesis of spherical silver nanoparticles with controllable sizes in aqueous solutions. *J Phys Chem C*.; 111(22):7910–7917.

- R. Nithya and R. Ragunathan, (2009). "Synthesis of silver nanoparticle using *Pleurotus sajor* caju, and its antimicrobial study," *Digest Journal of Nanomaterials and Biostructures*, vol. 4, no. 4, pp. 623–629.
- Rafieian-Kopaei M, Baradaran A. (2013). Combination of metformin with other antioxidants may increase its renoprotective efficacy. *J Renal Inj Prev.*; 2:35–6
- Rafieian-Kopaei M, Hosseini M, Shirzad H. (2014). Comment on: Effect of pomegranate flower extract oncis platin-induced nephrotoxicity in rats. *J Nephropathol.* ; 3:121–3.
- Reaven GM. (1995). The fourth Musketeer from Alexandre Dumas to Claude Bernard. Diabetologia; 38: 3-13
- Reilly, R.M. (2007). Carbon nanotubes: Potential benefits and risks of nanotechnology in nuclear medicine. J. Nucl. Med., 48, 1039–1042
- Rhead B, Karolchik D, Kuhn RM, Hinrichs AS, Zweig AS, Fujita PA, et al. The UCSC genome browser database: update 2010. Nucleic acids research. 2010;38(suppl_1):D613-D19.
- Richard J. Mahler, Michael L. (1 April 1999). Adler Type 2 Diabetes Mellitus: Update on Diagnosis, Pathophysiology, and Treatment. *The Journal of Clinical Endocrinology & Metabolism*, Volume 84, Issue 4, , Pages 1165–1171
- Rorbach-Dolata A, Kubis A, Piwowar A (2017) Epigenetic modifications: An important mechanism in diabetic disturbances. *Postepy Higieny i Medycyny Doswiadczalnej*; 71: 960-974.
- Rosenvinge A, Krogh-Madsen R, Baslund B, Pedersen BK. (2007). Insulin resistance in patients with rheumatoid arthritis: effect of anti-TNFalpha therapy. *Scand J Rheumatol.*; 36:91–6.

- Ruan H, Miles PD, Ladd CM et al. (2002). Profiling gene transcription in vivo reveals adipose tissue as an immediate target of tumor necrosis factor-alpha: implications for insulin resistance. *Diabetes*; 51:3176–88.
- S. Ahmed, M. Ahmad, B. L. Swami, and S. Ikram, (2016). "A review on plants extract mediated synthesis of silver nanoparticles for antimicrobial applications: a green expertise," *Journal of Advance Reseach*, vol. 7, no. 1, pp. 17–28.
- S.-H. Shin, M.-K. Ye, H.-S. Kim, and H.-S. Kang, (2007)."The effects of nano-silver on the proliferation and cytokine expression by peripheral blood mononuclear cells," *International Immunopharmacology*, vol. 7, no. 13, pp. 1813–1818.
- Saeedi P, Petersohn I, Salpea P, Malanda B, Karuranga S, Unwin N, Colagiuri S, Guariguata L, Motala A, Ogurtsova K, Shaw J, Bright D, Wiliams R, (2019) Global and regional diabetes prevelance estimates for 2019 and projections for 2030 and 2045: Results from International Diabetes Federation Diabetes Altas, 9th edition. *Diabetes Res Clin Pract*; Vol. 157
- Saratale, G.D., Saratale, R.G., Benelli, G. et al. (2017). Anti-diabetic Potential of Silver Nanoparticles Synthesized with Argyreia nervosa Leaf Extract High Synergistic Antibacterial Activity with Standard Antibiotics Against Foodborne Bacteria. J Clust Sci 28, 1709–1727.
- Saratale, R.G., H.S. Shin, G. Kumar, G. Benelli, D.-S. Kim and G.D. Saratale (2018). "Exploiting antidiabetic activity of silver nanoparticles synthesized using Punica granatum leaves and anticancer potential against human liver cancer cells (HepG2)." Artificial cells, *nanomedicine and biotechnology*; 46(1):211-222.
- Sarwar N, Gao P, Seshasai SR, Gobin R, Kaptoge S, Di Angelantonio et al. (2010). <u>Diabetes</u> mellitus, fasting blood glucose concentration, and risk of vascular disease: a

collaborative meta-analysis of 102 prospective studies. Emerging Risk Factors Collaboration. Lancet; 26;375:2215-2222.

- Scheen AJ. (2001). Obesity and diabetes. In : The management of obesity and related disorders (Kopelman P.G., Ed.), *Martin Dunitz Ltd, London, UK*: 11-44.
- Schulze MB, Manson JE, Ludwig DS. et al. (2004). Sugar-sweetened beverages, weight gain, and incidence of type II diabetes in young and middle-aged women. *JAMA*; 292:927–934.
- Sengottaiyan, A., Aravinthan, A., Sudhakar, C., Selvam, K., Srinivasan, P., Govarthanan,
 M., Manoharan, K., Selvankumar, T. (2016). Synthesis and characterization of
 Solanum nigrum-mediated silver nanoparticles and its protective effect on alloxaninduced diabetic rats. *J Nanostruct Chem* 6:41–48
- Shaheen T. I., El-Naggar M. E., Hussein J. S., et al. (2016). Antidiabetic assessment; in vivo study of gold and core-shell silver-gold nanoparticles on streptozotocin-induced diabetic rats. *Biomedicine & Pharmacotherapy*; 83:865–875.
- Sherin, A. (2015). National diabtes action plan of Pakistan: need and challenges. *KMUJ: Khyber Medical University Journal* 7(1):1-2
- Stensberg M.C., Wei Q., McLamore E.S., Porterfield D.M., Wei A., Sepúlveda M.S. (2011). Toxicological studies on silver nanoparticles: Challenges and opportunities in assessment, monitoring and imaging. *Nanomedicine*; 6:879–898.
- Stumvoll M, Goldstein BJ, van Haeften TW (2005). Type 2 diabetes: principles of pathogenesis and therapy. *The Lancet* 365: 1333-1346.

- Swarnalatha C, Rachela S, Ranjan P, Baradwaj P. (2012). Evaluation of *in vitro* antidiabetic activity of *Sphaeranthus amaranthoides* silver nanoparticles. *Int J Nanomat Biostr*; 2:25-29.
- Taghikhani A, Afrough H, Ansari-Samani R, Shahinfard N, Rafieian-Kopaei M. (2014). Assessing the toxic effects of hydroalcoholic extract of *Stachys lavandulifolia* Vahl on rat's liver. *Bratisl Lek Listy*; 115:121–4.
- Tamadon MR, Ardalan MR, Nasri H. (2013). World kidney day 2013; acute renal injury; a global health warning. *J Parathyr Dis.*; 1:27–8
- Tattersal RB, Fajans SS (1975) Prevalence of diabetes and glucose intolerance in 199 offspring of thirty-seven conjugal diabetic parents. *Diabetes* 24: 452-462.
- Tevatia, Siddharth. (2016). Nanotechnology: The Beginning of New Era. International Journal of Enhanced Research in Medicines & Dental Care ISSN: 2349-1590. 3. 7-14.
- Thatoi, P., Kerry, R. G., Gouda, S., Das, G., Pramanik, K., Thatoi, H., & Patra, J. K. (2016). Photo-mediated green synthesis of silver and zinc oxide nanoparticles using aqueous extracts of two mangrove plant species, Heritiera fomes and Sonneratia apetala and investigation of their biomedical applications. *Journal of Photochemistry and Photobiology B 163: 311-318*
- Thomas CC, Philipson LH (2015) Update on diabetes classification. *The Medical Clinics* of North America 99: 1-16.

Thomson PDR. (2004). PDR for Herbal Medicines. 3rd. Montvale, NJ, USA

Tian J, Wong KK, Ho CM, Lok CM, Yu WY, Che CM, *et al.* (2007). Tropical delivery of silver nanoparticles promotes wound healing. *Chem Med Chem*; 2:129-36.

- Tian, L.; Lu, L.; Qiao, Y.; Ravi, S.; Salatan, F.; Melancon, M.P. (2016). Stimuli-Responsive Gold Nanoparticles for Cancer Diagnosis and Therapy. *J. Funct. Biomater*, 7, 19.
- Tien DC, Liao CY, Huang JC, et al. (2008). Novel technique for preparing a nano-silver water suspension by the arc-discharge method. *Reviews on Advanced Materials Science*; 18:750–756.
- Tilg H, Moschen AR. (2008). Inflammatory mechanisms in the regulation of insulin resistance. *Mol Med.;* 14:222–31.
- Tripathi BK, Srivastava AK. (2006). Diabetes mellitus: complications and therapeutics. *Med Sci Monit.*; 12(7):RA130–147
- Tsuji T, Iryo K, Watanabe N, Tsuji M. (2002). Preparation of silver nanoparticles by laser ablation in solution: influence of laser wavelength on particle size. *Appl Surf Sci.*; 202(1–2):80–85.
- V. Thirumalai Arasu, D. Prabhu, and M. Soniya, (2010). "Stable silver nanoparticle synthesizing methods and its applications," *Journal of Biosciences Research*, vol. 1, no. 4, pp. 259–270.
- Vana D.R., Adapa D., VSS P., Choudhry A. (2019). Diabetes mellitus: Key genetic determinants and risk assessment. *Genet. Mol. Res*, 18(2)
- Wang, W.-X., B.-H. Wang, C.-Y. Yin, Y.-B. Xie and T. Zhou (2006). "Traffic dynamics based on local routing protocol on a scale-free network." *Physical review E* 73(2):026111
- Wichtl M. *Herbal Drugs and Phytopharmaceuticals*. Stuttgart, Germany: MedPharm Scientific Publishers; 2001

- Wing RR. (1995). Use of very low-calorie diet in the treatment of obese persons with noninsulin dependent diabetes mellitus. *J Am Diet Assoc*. 95:569–572
- Wu, Y., Ding, Y., Tanaka, Y., & Zhang, W. (2014). Risk factors contributing to type 2 diabetes and recent advances in the treatment and prevention. *International journal of medical sciences*, 11(11), 1185–1200.
- Yan Y, Sha Y, Yao G, Wang S, Kong F, Liu H et al. (2016). <u>Roux-en-Y Gastric Bypass</u> <u>Versus Medical Treatment for Type 2 Diabetes Mellitus in Obese Patients: A</u> <u>Systematic Review and Meta-Analysis of Randomized Controlled Trials</u>. *Medicine* (*Baltimore*); 95(17)
- Zhang J and Sun TQ (2009) IRS-2 and its involvement in diabetes and aging. *Vitamins and Hormones* 80: 389-407.
- Zimmet P, Alberti KG, Shaw J. (2001). Global and societal implications of the diabetes epidemic. *Nature*; 414(6865):782–787.
- Zoungas S, De Galan BE, Ninomiya T, Grobbee D, et al. (2009) Combined effects of routine blood pressure lowering and intensive glucose control on macrovascular and microvascular outcomes in patients with type 2 diabetes: New results from the ADVANCE trial. *Diabetes Care* 32: 2068-2074.
- Zullo A, Sommese L, Nicoletti G, Donatelli F, et al. (2017) Epigenetics and type 1 diabetes: mechanisms and translational applications. Translational Research: *The Journal of Laboratory and Clinical Medicine;* 185: 85-93.